

# Time for the end of GM/GE herbicide tolerant crops?



## EXECUTIVE SUMMARY

A report by GeneWatch UK

# Time for the end of GM/GE herbicide tolerant crops?

EXECUTIVE SUMMARY

A report by GeneWatch UK

August 2022

This is the Executive Summary of the full report, which is available on [www.genewatch.org](http://www.genewatch.org)



## GeneWatch UK

53, Milton Road, Cambridge, CB4 1XA, UK

Phone: +44 (0)330 0010507

Email: [mail@genewatch.org](mailto:mail@genewatch.org) Website: [www.genewatch.org](http://www.genewatch.org)

Registered in England and Wales Company Number 03556885

*Cover picture adapted detail from: String of monarch butterflies (Danaus plexippus) at Pismo Preserve in San Luis Obispo County, California USA, by Steve Corey at:*

<https://flickr.com/photos/22016744@N06/23289612339>

*This file is licensed under the Creative Commons Attribution-Share Alike 2.0 Generic license:*

## **Executive Summary**

In countries growing genetically modified (GM) crops, the adoption of GM crops which are tolerant to weedkillers is reaching saturation. These herbicide-tolerant (HT) GM crops have been genetically engineered so they can be blanket-sprayed with the associated herbicides, with the aim of killing weeds whilst the crop still grows. They were first grown commercially in 1996, when they were introduced by the US company Monsanto (now owned by Bayer). Monsanto's glyphosate-based weedkiller has the brand-name RoundUp, hence the first GM generation of herbicide-tolerant crops are tolerant to glyphosate and are known as 'RoundUp Ready' (RR) crops.

Herbicide-tolerant GM crops, which include an herbicide-tolerant trait alone or in combination with other traits, account for around 88% of the land area planted with GM crops worldwide (ISAAA, 2019a). Because cotton grown in India and China accounts for most non-HT GM crops, HT crops account for virtually all the GM crops grown for use in food or feed. This reality is in sharp contrast to GM industry PR which acts as a distraction by emphasising potential new traits, including GM crops that tolerate flooding or drought, which were first promised more than 40 years ago but have not been delivered. Since 1996, most herbicide-tolerant GM crops have been RR crops which are genetically engineered to be tolerant to glyphosate, but this has recently been changing with increasing areas planted with new HT crops which are tolerant to additional herbicides, such as dicamba and 2,4,D. These crops are mainly grown in North and South America, with the USA, Brazil and Argentina growing the largest quantities.

The aim of this report is to look at the economic, environmental and social impacts of growing RR crops and newer HT crops. This report reviews more than 25 years of experience with this technology.

We conclude that the cultivation of GM HT crops may be regarded as a temporary aberration, rather than the revolution originally proclaimed by the proponents of these crops. The growing failure of RoundUp Ready crops, due to the spread of glyphosate resistant (GR) weeds, provides an opportunity to phase out the use of RR crops and adopt new methods and technologies. The priority should be to reduce and replace the use of herbicides: not to replace RR crops with other herbicide-tolerant crops, whether or not these are GM crops or produced by different methods. It is particularly important that RR crops are not pushed into new countries which have so far avoided stepping onto the "transgenic treadmill", in which farmers are locked in to paying for ever more expensive seeds and herbicides. In seeking to expand markets for RR crops into new countries, the industry is dumping a failed technology on them.

### **Blanket spraying of RR crops with weedkiller leads to resistant weeds**

RR crops are designed to make farming practices easier in that they allow farmers to apply the weedkiller glyphosate during the cropping season without risking harm to their crops, which are genetically engineered to be tolerant to it. This has led to an unprecedented increase in the use of glyphosate (Powles & Preston, 2006; Duke & Powles, 2008; Grube et al 2011; Vivian et al. 2013; Benbrook, 2016; Myers et al., 2016).

Initial benefits for farmers adopting RR crops have vanished with the emergence of glyphosate resistant (GR) weeds, sometimes known as 'superweeds' (Vila-Aiub et al. 2007, 2008; Cerdeira et al. 2011; Benbrook, 2012a; Allison, 2015; Zhou et al., 2015;

Duke et al., 2018). Such weeds have evolved because RR crops allow farmers to blanket spray their crops with the weedkiller glyphosate instead of using a mix of approaches to tackling weeds, including crop rotation. Since GR weeds are no longer affected by spraying with glyphosate, they require the use of additional weedkillers or pulling up by hand (Caulcutt, 2009; Zhou et al., 2015). With 55 evolved GR weed species already known worldwide (Heap, 2021), and new GR weed species evolving at an increasing rate, RR technology is becoming obsolete. GR weeds are common in all RR crop producing countries and these are also the countries with the greatest area infested with GR weeds (Heap, 2021; Heap and Duke, 2018; Alcántara de la Cruz et al., 2020; Yanniccari et al., 2021; Pannell et al., 2017). RR crop cultivation is leading to increased herbicide application, thus adding costs for farmers, and increasing risks to the environment and human health. On the two most important GM crops in the US, corn and soybean, the total applied toxicity of pesticides (not just glyphosate) has increased along with increasing GM adoption, notably since 2008 as GR weeds became a greater problem (Schulz et al., 2021).

In response to the problem of glyphosate resistant weeds, the industry has developed new GM herbicide tolerant crops which are resistant to additional weedkillers, such as 2,4-D and dicamba, as well as glyphosate. Such crops exacerbate concerns about adverse environmental impacts, pesticide residues in the food chain, and the future evolution of weeds which will become resistant to multiple herbicides (Mortensen et al. 2012, Roseboro, 2012).

### **Patents and monopolies add further costs and prevent seed saving**

Just four large firms (Bayer, Corteva, ChemChina-Syngenta, and BASF) control around 70% of the global pesticides market and 60% of the global seed market (Clapp, 2021). Patents on GM seeds give companies monopoly control and allow them to prevent seed saving. This, along with market concentration in the industry, has led to significant increases in seed prices, with high premiums for GM seeds and restrictions on the non-GM varieties available on the market in some countries (Mascarenhas & Busch, 2006; Howard, 2009; Zilberman et al. 2010; Benbrook, 2012a; Filomeno, 2013; Benbrook, 2018; Brunharo et al., 2022). Farmers buying RR seeds are locked into a “transgenic treadmill” in which they are forced to pay for hikes in seed prices and for increasing amounts of herbicides and labour to tackle weed resistance (Binimelis et al., 2009; Mortensen et al., 2012).

### **RoundUp Ready crops do not have higher yields**

Most farmers growing RR crops have adopted RR crop technology in the hopes of increasing their yields (Fernandez-Cornejo et al., 2014). However, there are no RR crops available today that increase the yield potential of a hybrid variety: any benefit to yields arises only if these crops improve weed control (Gurian-Sherman, 2009; Nolan & Santos, 2012; Bruns, 2014). A global meta-analysis of studies by Areal et al. (2013) reports no significant differences in yields between RR and conventional crops. Yield data from North America and Western Europe shows that Western Europe, where to date no herbicide tolerant resistant crops are grown, had a greater yield increase between 1961 and 2010 than North America for oil seed rape and maize (which are predominantly RR crops in North America) and that overall yields were similar or higher in Europe than in the USA (Hilbeck et al. 2013; Heinemann et al., 2014a, 2014b). There is also some evidence that suppressed plant defence and enhanced disease susceptibility caused by glyphosate may have a negative impact on RR plants, through adverse effects on beneficial soil micro-organisms and plant nutrient uptake (Sanogo et al., 2000; King et al., 2001; Eker et al., 2006; Bellaloui et

al. 2008; Bott et al., 2008; Johal and Huber, 2009; Zobiole et al., 2010a&b, 2011, 2012; Freitas-Silva et al., 2021).

### **Demand for non-GM seeds and ingredients is increasing**

Farmers worldwide also need to consider the demand for non-GM ingredients, which is forecast to grow significantly (Mordor Intelligence, n.d.; Grand View Research, 2019; Fortune Business Insights, 2022).

Demand for animal feed that is segregated as non-GM has grown, particularly in Europe (Tillie & Rodríguez-Cerezo, 2015), where price premiums for non-GM crops reflect the preference of European consumers for non-GM products (Gaitán-Cremaschi et al., 2015; Fortune Business Insights, 2022). By 2021, around 60-70% of all milk egg, poultry and meat production in Germany was certified according to the GM-free VLOG standard (Southey, 2021). Brazil has increased its import share faster in countries with a strong non-GM preference versus other countries. This is explained statistically by Brazil's level of non-GM soybean production rather than by changes in prices. Garrett et al. (2013) find that the Netherlands, Italy, Spain, and Belgium increased imports from Brazil and simultaneously decreased imports from the United States, even as Brazil's currency increased in value in the late 2000s, which should have made Brazilian soybean producers less competitive than their North American counterparts on a pure cost basis. The South American non-GMO food market continues to grow, with Brazil expected to have the biggest market (Fortune Business Insights, 2022). Among US farmers, interest in growing non-GM varieties reportedly started increasing around 2014, with seed companies reporting strong demand for non-GM seed sales and some even reporting they had sold out of non-GM seeds due to the rapidly increasing demand (Bunge, 2015; Roseboro, 2015b; Doering, 2015; Kuphal, 2017). Retail sales of products verified by the Non-GMO Project, based in North America, rose dramatically from \$248.8 million in 2010 to \$8.5 billion in 2014 and 13.5 billion in 2015, with sales now over \$26 billion (Non-GMO Project, 2014; 2015; 2022). However, the non-GM corn and soybean supply in the U.S. remains relatively small (Twellman, 2021).

### **Growing GM crops risks expensive contamination incidents**

Cultivation of RR crops risks GM contamination of non-GM food and feed supplies (Price & Cotter, 2014). Contamination risks arise due to cross-pollination of non-GM crops and co-mingling of seeds or grains during harvest, transportation, storage, processing and distribution (Sohn et al., 2021). These risks cannot be eliminated through technical measures (Binimelis, 2008; Paull, 2018; Lu et al., 2019) and this causes legal and economic uncertainties for farmers, because contaminated crops have lower value (due to consumers' preference for non-GM crops) and may be rejected completely by some markets (e.g. organic markets, or any market where the GM crop has not been authorised by regulators).

Attempts to allow GM and non-GM crops to be grown together in a given country or region (known as "co-existence") creates tensions among neighbouring farmers because of the risk of GM contamination. Every actor and level of a supply chain will be economically affected under a coexistence scenario and costs of coexistence of GM and non-GM agricultural production systems are influenced by multiple factors (Gabriel & Menrad, 2015). At the producer level they include costs for cleaning of machinery and equipment, buffer zones of uncultivated land around the edge of non-GM fields, monitoring costs such as testing of seeds or crops and building additional farm storage facilities. For processors, costs to prevent contamination include: costs for testing of the incoming commodity as well as the produced outgoing goods,

greater transportation distances to the next GM or non-GM plant respectively, building of additional storage facilities, complete second production line in an existing plant, cleaning or flushing of repositories, investment in additional personnel and equipment and in training programs for workers (Gabriel & Menrad, 2015). According to this study, the total additional costs of coexistence and implemented product segregation systems can amount up to 14% of the total product turnover at the gates of rapeseed oil mills or companies processing maize starch, respectively. In Switzerland, where GM crops are not grown, estimated coexistence measures if they were introduced could amount to up to 5-20% of the total costs for conventional production (Albisser Vögeli et al., 2011). The costs to prevent GM contamination are likely to be especially high for organic producers, since global organic farming standards do not allow GMOs in either seed or food (IFOAM, 2002).

Thus, allowing GM cultivation increases the cost of food supplies, because of the added costs of segregation. In countries where GM crops are grown, non-GM farmers, including organic farmers, bear risks and costs associated with protecting their crops from GM contamination and certifying their supply chain as GM-free for consumers.

When coexistence measures fail, contamination incidents can lead to the destruction of crops or entire fields (Furst, 1999; Smyth et al., 2002) and the rejection of shipments, product recalls and loss of markets (Ryan & Smyth, 2012; Smyth et al., 2002; Schaefer & Carter, 2015; USDA NASS, 2015; Reuters, 2016c), with multi-million dollar economic impacts.

GM contamination can also have environmental implications and risk the loss of local varieties of seed. Glyphosate tolerance, and other GM traits, can spread from GM maize to maize landraces, as has happened in Brazil (Fernandes et al., 2022). Maize is mainly produced by smallholders in Mexico, using landraces that are very well adapted to the local growth conditions. Contamination of these landraces, could threaten preservation of this very important maize genetic diversity (Snow, 2009). GM transgenes are already present in at least some maize landraces in Mexico (Piñeyro-Nelson et al., 2009; Quist & Chapela, 2001; Snow, 2009). Wild populations of the most widely cultivated cotton species in the world, *Gossypium hirsutum*, have also been contaminated by GM varieties, the majority of which are geographically located over 300 km away from all wild cotton populations (Wegier et al., 2011). In Spain, GM contamination of organic maize led to the loss of farmers' maize varieties adapted to the local climate (Cipriano et al., 2006). Such events could limit the future availability of high-value germplasm in breeding programs (Burgeff et al., 2014).

### **Impact of RR crops on farmers' choice, land rights and indebtedness**

Patents on GM crops lead to restricted access to breeding material for farmers and breeders and thus hinder innovation in plant breeding and impede farmers' freedom of choice. In countries adopting GM crops the maize seed market is more concentrated with fewer available maize cultivars for farmers than in non-adopting countries, where it has become more difficult to find non-GM seeds (Roseboro, 2008; Hilbeck et al., 2013; Burgeff et al., 2014). In the USA, rising input costs, volatile production values, and rising land rents have left farmers with unprecedented levels of farm debt, low on-farm incomes, and high reliance on federal programs (Burchfield et al., 2022). Subsidies are largely directed at commodity production, including soy and corn, which are typically GM crops, and for which per acre costs tripled between 1990 and 2020.

Impacts of RR crop cultivation on smallholders in South America and elsewhere include land conflicts and the intensification of agro-industrial practices, including greater use of herbicides, increased farm sizes, land use changes and deforestation, seed price hikes, and the expansion of monocultures and indebtedness (Lapegna, 2013; Garrett and Rausch, 2016; Goldfarb and van der Haar, 2016; Leguizamón, 2016; McKay and Colque, 2016; Elgert, 2016; MASIPAG, 2013; Phélinas and Choumert, 2017; Schmidt et al., 2022; Dreoni et al., 2022).

### **RR crops have negative environmental impacts**

The widespread adoption of RR crops in North and South America has contributed significantly to an increased environmental presence of glyphosate-based herbicides and their primary break-down product, AMPA, in rain, streams, rivers, lakes, ponds, wetlands, soil water, ground water, plants, soil, dust and sediment (Battaglin et al. 2005, 2014; Struger et al. 2008; Chang et al., 2011; Bohm et al., 2014; Majewski et al., 2014; Bento et al., 2016; Mamy et al., 2016; Bonansea et al., 2017; Alonso et al., 2018; Fernandes et al., 2019; Zheng et al. 2018; Clasen et al. 2019; Iturburu et al., 2019; Lupi et al., 2019; Lutri et al., 2020; Maggi et al., 2020; Medalie et al., 2019; Montiel-León et al., 2019; da Silva et al., 2021; Barbosa Lima et al., 2021; Botten et al., 2021; Brovini et al. 2021a,b; Cristofaro et al., 2021; Ramirez Haberkon et al., 2021; Carretta et al., 2022).

Negative environmental impacts due to growing herbicide-tolerant GM crops, including RR crops, include:

- impacts on farmland diversity of weeds, insects and birds through loss of important habitats due to blanket spraying of these crops with herbicide (Burke, 2003; Burke, 2005; Firbank et al., 2003; Gibbons et al., 2006; Cederlund, 2017; Pereira et al., 2018a, 2020);
- chronic toxicological effects of glyphosate and its metabolites on annelids (earthworms), arthropods (crustaceans and insects), molluscs, echinoderms, fish, reptiles, amphibians, birds, mammals, and non-target plants (Santadino et al., 2014; Zaller et al. 2014; Gaupp-Berghausen et al. 2015; Domínguez et al., 2016; Kissane & Shephard, 2017; Gill et al., 2018; Odetti et al., 2020; Ruuskanen et al., 2020a,b,c; Singh et al., 2020; Barbosa Lima et al. 2021);
- negative effects on pollinators, such as bees, including damage to habitat and ecosystems; toxicity; and effects on their behaviours, growth and development, metabolic processes, and immune defence (Fuchs et al., 2021; Strandberg et al., 2021; Battisti et al., 2021; Tan et al., 2022);
- toxic and chronic sub-lethal effects of glyphosate-based weedkillers on aquatic species including tadpoles, frogs, snails, crayfish, molluscs, crabs, fish, fresh-water fleas and corals (Relyea, 2005a,b and c; Pérez et al., 2012; Cuhra et al., 2013, 2014, 2015; Avigliano et al. 2014a,b; Gonçalves et al., 2019; Hendlin et al., 2020; Herek et al., 2021; Matozzo et al. 2020; Mohapatra et al., 2021; Moutinho et al., 2020; Riaño et al. 2020; Slaby et al., 2020; Suppa et al., 2020; Babalola et al., 2021; Le Du-Carrée et al., 2021, 2022; Ramsdorf et al., 2021; Rodríguez et al., 2021; Sánchez et al., 2021; Santos-Silva et al., 2021; Tresnakova et al., 2021; de Maria et al., 2021, 2022; Jia et al. 2022; Liu et al., 2022b; Pompermaier et al., 2022; Zhou et al., 2022); and
- adverse impacts of glyphosate-based herbicides on soil biota: such as effects on soil microbial communities (Jaworski, 1972; Schulz et al., 1985; Moorman et al., 1992; Dick and Quinn, 1995; Kremer and Means 2009; Nye et al., 2014; Newman et al., 2016); and impacts on overall ecosystem functioning, including interactions of crops with fungi and soil-borne pathogens (Johal &

Rahe 1984; Sanogo et al., 2000, 2001; Larson et al., 2006; Krzysko-Lupicka and Sudol, 2008; Johal and Huber, 2009; Kremer and Means, 2009; Zobiole et al., 2011; Lu et al., 2018; Martinez et al., 2018; Yang et al., 2020; Hertel et al. 2021, Van Bruggen et al, 2021; Vázquez et al., 2021; Chávez-Ortiz et al., 2022).

One important example of the effects of habitat loss is a major contribution to the dramatic decline in populations of the Monarch butterfly in the USA. Although other factors (such as climate change and deforestation) play a role, this decline is associated with the loss of the milkweed habitat where the butterflies lay their eggs, caused by blanket spraying the weedkiller glyphosate on RR crops (Hartzler, 2010; Zalucki & Lammers 2010; Brower et al., 2012; Pleasants & Oberhauser, 2012; Fallon, 2014; Flockhart et al., 2014; Vidal & Rendón-Salinas, 2014; Stenoien et al., 2016; Pleasants, 2017; Pleasants et al., 2016, 2017; Thogmartin et al., 2017; Belsky & Joshi, 2018; Malcolm, 2018; Taylor et al., 2020).

Glyphosate-contaminated runoff also likely contributes to harmful incidences of algal bloom in lakes (Dabney & Patiño, 2018; Berman et al., 2020).

In glyphosate-based herbicide formulations, glyphosate is the active ingredient that is supposed to kill the target weeds. Those formulations also contain various adjuvants, the so-called inert ingredients, including surfactants such as polyethoxylated tallow amine (POEA) which is found in Roundup. However, ecotoxicological assessment of pesticides usually focuses on the effects of the active ingredient, such as glyphosate, rather than on commercial formulations like Roundup (Cox & Sorgan, 2006; Pereira et al., 2009; Mesnage & Antoniou, 2018; Sprinkle & Payne-Sturges, 2021; Martins-Gomes et al., 2022). This is a major issue of concern because many studies find that commercial formulations are significantly more toxic than glyphosate alone, particularly to aquatic organisms (Mitchell et al., 1987; Servizi et al. 1987; Mann & Bidwell, 1999; Perkins et al. 2000; Marc et al., 2002; Everett & Dickerson, 2003; Tsui & Chu, 2003, 2004; Howe et al., 2004; Cedergreen & Streibig, 2005; Brausch et al. 2007; Brausch & Smith, 2007; Pereira et al. 2009; Moore et al. 2012; Vincent & Davidson, 2015; Bach et al., 2016; Rissoli et al., 2016; Janssens & Stoks, 2017; de Brito Rodrigues et al., 2019; Mesnage et al. 2019; Bednářová et al., 2020; Le Du-Carrée et al., 2022; Sabio y García et al., 2022).

In South America, there have also been significant changes in land use to create large-scale RR soybean farms, for example in the Rolling Pampas in Argentina and the Cerrado in Brazil, with serious negative impacts on biodiversity and water-balance (De la Fuente et al. 2006, 2010; Martinelli et al., 2010; Hayhoe et al., 2011; Macedo et al., 2013; Neill et al., 2013; Redo et al. 2013; Eloy et al. 2016; de Groot et al., 2021).

An additional issue with RR crops is that they may contribute to the development and spread of antibiotic resistant bacteria, which can make it difficult to treat human and animal bacterial infections. Some RR crops contain antibiotic resistant marker genes, which might be able to spread into the environment (Chen et al., 2012). Exposure to sub-lethal levels of the herbicide Roundup has been linked to a change in susceptibility of bacteria to antibiotics, significantly increasing the concentration of two antibiotics (kanamycin and ciprofloxacin) necessary to kill gut bacteria associated with food poisoning, *Escherichia coli* and *Salmonella enterica* (Kurenbach et al., 2015). This research suggests that spraying RR crops with RoundUp might contribute to the development of antibiotic resistant bacteria in the environment, with major implications for human and animal health (Van Bruggen et al., 2018; Raoult et al., 2021; Liao et al., 2021; da Costa et al., 2021; Daisley et al., 2022).

## **RR crops pose unknown risks to human health**

There are significantly higher levels of glyphosate and AMPA residues in RR soybeans compared to conventionally grown or organic soybeans (Arregui et al., 2004; Bøhn et al., 2014; Bohm et al., 2014). Bøhn & Millstone (2019) estimate that glyphosate-tolerant soybeans produced on commercial farms in the USA, Brazil and Argentina accumulate in total an estimated 2,500–10,000 metric tonnes of glyphosate per year, which enter global food chains. Glyphosate has been detected in a wide variety of foods, including soy-based infant formula and honey: dietary exposure levels are generally (but not always) below permitted limits (Rodrigues & de Souza, 2018; Bøhn & Millstone, 2019; Xu et al., 2019; de Souza et al., 2021; Rodrigues et al., 2020; Louie et al., 2021; Viljoen et al., 2021). However, regulatory limits vary in different countries, there is a lack of transparency about how they are set, and some researchers believe that the risks to human health could still be underestimated (Marino et al., 2021).

Krüger et al. (2014a) showed that glyphosate that accumulates in feed can be consumed by animals and be detected in their organs and urine. Subsequently, glyphosate has been detected in the urine of adults and children, both within and outside agricultural communities (Gillezeau et al., 2020; Ferreira et al., 2021; Lozano-Kasten et al., 2021; Grau et al., 2022; Nomura et al., 2022). Farmers and other operators can be directly exposed to glyphosate-based formulations when they are spraying it onto their fields (Acquavella et al., 2004; Mesnage et al., 2012). Children may also be exposed to glyphosate-contaminated breast milk. Glyphosate was detected in all breast milk samples taken from mothers in a study in Brazil, undertaken at the peak of glyphosate application in corn and soy crops (Camiccia et al., 2022). Regulators do not currently routinely monitor levels of glyphosate in food and have not investigated reports that glyphosate may be detected in human urine samples and breast milk as a result of its presence in the food chain. Some studies suggest that spraying with glyphosate-based weedkillers may also adversely affect the nutrient composition of soybeans (Zobiolo et al., 2010b,c; Bellaloui et al., 2008).

As discussed above, Roundup formulations are a mixture of glyphosate and other chemicals that have been shown to increase the toxicity of glyphosate to aquatic organisms. Many toxicological studies conducted with human, mouse and rat cells confirm these findings and suggest that looking at the effects of glyphosate alone is insufficient for a comprehensive assessment of the possible risks to human health resulting from growing and consuming RR crops (Benachour et al., 2007; Benachour & Séralini, 2009; Clair et al., 2012; Gasnier et al., 2009; Mesnage et al., 2013, 2014; Moore et al., 2012; Richard et al., 2005; Walsh et al., 2000; Young et al., 2015; Chłopecka et al., 2017; Vanlaeys et al., 2018; Dedeker et al., 2018; Defarge et al., 2018). However, regulators only consider the effects of glyphosate alone (Mesnage et al., 2019).

In March 2015, the World Health Organisation (WHO)'s cancer agency, the International Agency for Research on Cancer (IARC), classified glyphosate as "probably carcinogenic to humans" (Guyton et al., 2015). As a consequence, many countries and regions have restricted the use of glyphosate (Where is Glyphosate Banned?, 2022). Subsequent reviews of the evidence have confirmed that chronic exposure to glyphosate causes a variety of tumours in rats and mice, and that there is clear evidence of glyphosate toxicity in studies using human cells (Agostini et al., 2020; Portier, 2020).

Research in Sri Lanka and elsewhere suggests a possible link between simultaneous exposure to glyphosate and toxic heavy metals, and chronic kidney disease, with other factors (such as exposure to high temperatures and other pollutants) perhaps playing a role (Jayasumana et al., 2014, 2015; Gunatilake et al., 2019; Herrera-Valdés et al., 2019; Babich et al., 2020; Abdul et al., 2021; Upamalika et al., 2022).

There is evidence that glyphosate may act as an endocrine disrupting chemical (EDC) i.e. a chemical that interferes with female and male sex hormones (Richard et al., 2005; Benachour et al., 2007; Gasnier et al., 2009; Romano et al., 2010; Clair et al., 2012; Thongprakaisang et al., 2013; Abarikwu et al., 2015; Guerrero Schimpf et al., 2017; Varayoud et al., 2017; Anifandis et al., 2017, 2018; Cai et al., 2017; Ingaramo et al., 2017, 2020, 2022; Owagboriaye et al., 2017; Lorenz et al., 2020; Kaboli Kafshgiri et al., 2021; Lesseur et al., 2021; Milesi et al., 2021; Mohammadi et al., 2021; Muñoz et al., 2021; Serra et al., 2021; Zhao et al., 2021). Endocrine disruptors can lead to negative impacts on male and female reproductive health, even at very low doses. These effects are not adequately regulated (Kalofiri et al., 2021). Two small studies have found that glyphosate exposure (measured in urine) in pregnancy is correlated with shortened pregnancy lengths (Parvez et al., 2018; Silver et al., 2021).

Other researchers suggest that glyphosate could affect gut bacteria, killing beneficial bacteria and allowing harmful ones to cause disease (Krüger et al., 2013; Shehata et al., 2013; Pu et al., 2020, 2021; Barnett et al., 2022).

Working with glyphosate and glyphosate spray drift can affect farm workers, bystanders and people living in the surrounding area.

In the Ontario Farm Family Health Study, Arbuckle et al. (2001) observe moderate increases in risk of early abortions for preconception exposures to any herbicide, and for late abortions, preconception exposure to glyphosate is associated with elevated risk. In the same study, Savitz et al. (1997) find that combinations of farm activities using a variety of chemicals, including glyphosate, are associated with an increased risk of miscarriage in the wives of exposed farm workers. In the Red River Valley, Minnesota, USA, Garry et al. (2002) find that exposure to glyphosate is associated with an increased risk of neurobehavioral developmental effects. In the Agricultural Health Study in Iowa and North Carolina, Hoppin et al. (2008) find an increased risk of atopic asthma in farm women using glyphosate and a number of other pesticides, and Hoppin et al. (2016) find an increased risk of allergic and non-allergic wheeze in male farm workers using glyphosate and some other pesticides. In the same study, Slager et al. (2009) find an increased risk of rhinitis in farm workers who had used glyphosate in the past year.

Aerial application (spraying from planes) increases the risk of accidental exposure of neighbouring inhabitants (Schiesari and Grillitsch, 2010; Pignati et al., 2007). Epidemiological studies and reports of interviews with local people cannot prove cause and effect, nevertheless there are numerous and widespread reports of glyphosate poisonings due to aerial spraying of RR soybeans in Latin America (Benítez-Leite et al., 2007; Oliva et al., 2008; Berger and Ortega, 2010; Sineiro and Berger 2012; Rigotto et al., 2014; Oliveira et al., 2014; Silva et al., 2015; Elgert, 2016; Lapegna, 2016; Dias et al., 2020; Longhi & Bianchi, 2020). Reported effects, according to people living in sprayed areas, include vomiting, diarrhoea, respiratory problems, skin rashes, cancer, infertility, pregnancy problems, and birth defects (PAN Asia & Pacific, 2008 & 2012).

## **RR crops do not help to feed the world or tackle climate change**

The primary reasons for hunger are poverty and lack of access to affordable food (Tscharntke et al., 2012). Conflict, weather extremes and economic shocks were the main drivers behind food insecurity in 2021, with poverty and inequality as underlying causes (EU/FAO/WFP, 2022).

RR crops are currently produced mainly for use in animal feed (soya and maize/corn) or in biofuels (corn ethanol) or fabric (cotton). Soybean and maize (corn) are the top two GM crops grown by area, the majority of which are Roundup Ready. Most soy (around 75% measured by weight in 2018) is fed to animals in livestock production systems, with around 3.8% going to biofuels and other industrial applications, and only 19.2% to direct human consumption as food (mainly as soybean oil) (Fraanje & Garnett, 2020). Similarly, around 74% of the global maize production is used for animal feed (Cassidy et al., 2013). In the U.S., 40% of the maize harvest was processed to ethanol in 2014 (Ranum et al., 2014). In 2014, production and use of corn ethanol resulted in 27 billion kg more carbon emissions than if conventional gasoline were used according to calculations by the Environmental Working Group (Cassidy, 2015a). This is because converting rainforests, peatlands, savannas, or grasslands to produce food crop-based biofuels in Brazil, Southeast Asia, and the United States releases 17 to 420 times more carbon dioxide than the annual greenhouse gas (GHG) reductions that these biofuels would provide by displacing fossil fuels (Fargione et al., 2008).

Shifting crop calories used for animal feed and biofuels to direct human consumption could, according to Cassidy et al. (2013), potentially feed an additional 4 billion people and in the U.S. alone an additional 1 billion people. Further, tackling food waste can also play a major role: many crop calories are lost during food production, transport and storage as well as in retail facilities, restaurants and at private households etc. (FAO, 2011).

It is questionable whether sparing land for nature needs higher intensity of farming to produce adequate food (Tscharntke et al., 2012). Strategies to increase yields without explicitly considering the actual and potential cost of biodiversity losses can compromise ecosystem functionality and resilience in agriculture. Rather, food security and food sovereignty need to increase in areas where the hungry live, based on robust, eco-efficient approaches. Smarter resource use, improving livelihoods of small-scale farmers, reducing food waste and small changes in diets, such as reducing meat consumption or swapping from grain-fed beef to chicken or grass-fed beef, have the potential to double calorie availability (Cassidy 2015b).

Further, in the case of RR crops, yields have not increased compared to non-GM crops (Areal et al., 2013). Cultivation of RR crops has led to significant expansion of intensive agricultural monocultures into previously diverse ecosystems (Oliveira and Hecht, 2016) and production of non-GM soybean meal has been found to be more sustainable than GM soy production (Ortega et al. 2005; Gaitán-Cremaschi et al., 2015).

Some authors have argued that the use of no-till agriculture (i.e. farming without disturbing the soil through ploughing), in combination with RR crops, has helped to mitigate climate change by keeping carbon in the soil: however, in a 41 year experiment in France, no-till agriculture led to no increase in soil organic carbon (Powlson et al., 2014). In addition, whilst the use of no-till increased in the United States from 1998 to 2016, it then shrank again, although herbicide-tolerant GM corn and soybeans still dominate the market (Yu et al., 2020). This is likely at least partly

due to the increasing presence of glyphosate-resistant weeds, which have led to a return to ploughing.

### **Industry responses to glyphosate-resistant weeds are not sustainable**

The industry's answer to the development of GR weeds is mainly herbicide-centric and includes a) developing herbicide tolerant (HT) crops with enhanced tolerance to glyphosate (allowing higher application rates), b) increasing the herbicide platform used on RR crops to include additional herbicides (e.g. in seed treatments and tank mixes); and c) developing new HT crops with tolerance to additional herbicides (Desquilbet et al. 2019).

Another aspect of the industry response is the use of other (supposedly beneficial, but likely ineffective) traits as a 'Trojan Horse' to smuggle herbicide tolerant GM traits into new crops and markets. These include HB4 GM wheat, developed by Bioceres, which is tolerant to glufosinate, but is being promoted for its supposedly drought tolerant properties (Paixão, 2020; Little, 2022); camelina (a plant also known as 'false flax') with herbicide-tolerance combined with altered oil content (Yield10 Bioscience, 2022, ACRE, 2019); and drought-tolerant GM maize for Africa, which is also being stacked with glyphosate-tolerance in some cases (African Centre for Biodiversity, 2021). These projects are consistent with the industry's awareness that, although RoundUp Ready crops are failing, there may still be opportunities to profit from expanding into new geographic areas and/or new crops before resistant weeds take hold (Green & Siehl, 2021). This PR strategy acts as a distraction from the negative consequences of growing HT GM crops, and as a means to attempt dump failing HT traits onto new markets.

Increased spraying of tank mixes of multiple weedkillers has led to grower weed control costs tripling in the USA (Vivian et al, 2013; Evans et al. 2016; Myers et al., 2016; Pratt, 2016a; Duke et al., 2018). The total applied toxicity of pesticides (not just glyphosate) has increased significantly since 2008 (Schulz et al., 2021). A 2015 survey conducted in 17 states in Brazil, revealed that 97% of respondents used tank mixtures by this date, usually with 2 to 5 products at the highest recommended doses (Gazziero 2015). A major problem remains the inadequate examination by regulators of the effects of mixtures of herbicides on human health and the environment (Sprinkle & Payne-Sturges, 2021). In addition, weeds are becoming resistant to multiple different herbicides (e.g. Benoit et al., 2020).

The US Department of Agriculture (USDA) argues that GM maize and soybeans with resistance to multiple herbicides will become the norm in future (Nandula, 2019). GM soybeans and maize with resistance to dicamba and 2,4-D are already on the market, and these are being stacked with existing GM resistance traits (to glyphosate and/or glufosinate) or other herbicides (such as isoxaflutole). In the USA, in crop year 2018, around three quarters of the soybean seed offered to farmers expressed the glyphosate-resistance gene, plus either dicamba or 2,4-D resistance genes (Benbrook, 2018). In 2019, Monsanto (now owned by Bayer) filed a petition with the USDA for determination of nonregulated status of a genetically engineered corn variety resistant to five active ingredients: glyphosate, glufosinate, dicamba, 2,4-D and quizalofop (Monsanto, 2019). These herbicide tolerant GM crops allow farmers to apply additional herbicides such as 2,4-D, dicamba, isoxaflutole or glufosinate during the whole cropping season at high rates, with the risk of detrimental effects to the environment and human health. For example:

- isoxaflutole is known to persist in the environment and to leach into and accumulate in ground- and surface waters (US EPA, 1998);

- an association between increasing 2,4-D application and human urine concentrations has already been reported (Freisthler et al., 2022); 2,4-D is classified as possibly carcinogenic by the WHO (IARC, 2018); 2,4-D is reported to be toxic to a variety of organisms, including fish, amphibians, insects, earthworms and rodents (Islam et al., 2018; da Silva et al., 2022);
- dicamba is a suspected endocrine disruptor (Zhu et al. 2015); and
- glufosinate is classified as a known or presumed reproductive toxicant and is no longer authorised for use in the EU (EFSA, 2017; European Commission, n.d.).

Moreover, 2,4-D and dicamba are prone to drift (risking damage to other farmers' crops, as well as the environment) (Murschell & Farmer, 2019; Lerro et al., 2020; Soltani et al., 2020) and it has been shown that repeated herbicide drift exposure can rapidly select for weed resistance (Vieira et al. 2020; Comont et al., 2020). Dicamba-resistant, 2,4-D resistant and glufosinate-resistant Palmer Amaranth (pigweed) have already been identified in the USA (Kumar et al, 2019; Unglesbee, 2020b; Unglesbee, 2021a). This circular process of the evolution of resistant weeds and the subsequent development of the next generation of transgenic crops, that allow for an intensified use of herbicides and thus favour the emergence of another round of resistant weeds, has been called the "transgenic treadmill" (Binimelis et al., 2009; Mortensen et al., 2012).

RR GM crops, and newer HT GM crops, use a method of genetic engineering known as transgenesis, which involves transferring new DNA from another species into plant cells (known as 'transgenes'). Newer genetic engineering techniques, using a variety of methods called 'gene editing', may allow new herbicide-tolerant GM crops to be produced which rely on mutating the crop's own genes and not on introducing foreign genes into the genome of a crop. There is commercial interest in this approach because such crops may be deregulated in some countries, so that environmental risk assessments and food labelling may not be required before they can be marketed. In particular, 'base editing' and 'prime editing' techniques can be used to mutate DNA without the need for donor DNA, although these methods are not currently efficient (Tang et al., 2020). Crops that have been gene edited to include herbicide tolerant traits remain at the experimental stage, but include wheat, rice, maize, soybean, potato, rapeseed (canola), flax, cassava, watermelon and tomato (Gosavi et al., 2022). The problems associated with existing HT GM crops will not be avoided by using gene editing techniques, since all these experimental crops are genetically engineered to withstand blanket spraying with the associated herbicides.

## **Lawsuits**

There have been numerous lawsuits relevant to the cultivation of GM herbicide-tolerant crops in the United States.

One set of lawsuits relates to claims that exposure to glyphosate causes cancer and environmental harm. Following the IARC's classification of glyphosate as a "probable human carcinogen", in March 2015, numerous lawsuits were filed alleging that past use of Monsanto's Roundup herbicide had contributed to the plaintiffs' development of non-Hodgkin lymphoma (NHL). Three lawsuits were heard before a jury and resulted in victories for the plaintiffs. In June 2022, the U.S. Supreme Court rejected Bayer's bid to dismiss these legal claims by customers and left in place the lower court decision that upheld \$25 million in damages awarded to one California resident (Hurley, 2022). In July 2021, Bayer (which bought Monsanto in 2018) took an additional litigation provision of \$4.5 billion for this case, on top of \$11.6 billion that

the company previously set aside for settlements and litigation (Hurley, 2022). In addition, in a case brought by farmers and environmental groups, the 9th Circuit Court of Appeals in California determined in June 2022 that the EPA did not adequately consider whether glyphosate causes cancer and threatens endangered species, and ordered it to look again at the risks it poses (Stempel, 2022).

Another set of lawsuits relates to crop damage caused by farmers spraying dicamba or 2,4-D on to GM crops resistant to these herbicides. In particular, Elmore (2022) describes how, in 2021, thousands of U.S. growers reported to the Environmental Protection Agency (EPA) that dicamba sprayed by other farmers on dicamba-resistant GM crops damaged crops in fields all over the country. In February 2020, Bader Farms won the first dicamba lawsuit and was awarded U.S. \$15 million in damages, plus U.S. \$250 million in punitive damages. The jury also found Monsanto and BASF had engaged in a joint venture and conspiracy, knowingly risking widespread crop damage in order to increase their own profits (Davies, 2020; Gillam 2020b). In June 2020, the Ninth Circuit's three-judge panel unanimously vacated EPA's approval of dicamba based herbicides (National Family Farm Coalition v. USEPA, 2020; Unglesbee, 2020f). However, these were subsequently re-registered. In 2022, the federal judge considering a case against the EPA in the U.S. District Court for the District of Arizona ordered that the EPA should file a report on the status of its ongoing evaluation of its options for addressing future dicamba-related incidents (Unglesbee, 2022a). The environmental and farming organisations involved subsequently asked court to lift a stay and expedite their lawsuit demanding EPA vacate its 2020 dicamba herbicide registrations (Unglesbee, 2022b). Despite these developments, the companies involved aim to commercialise new dicamba tolerant traits, some with tolerances against four or five active ingredients (Unglesbee, 2020n).

## Alternatives

The growing failure of RoundUp Ready crops, due to the spread of glyphosate resistant (GR) weeds, provides an opportunity to phase out the use of RR crops and adopt new methods and technologies. The priority should be to reduce and replace the use of herbicides: not to replace RR crops with other herbicide-tolerant crops, whether or not these are GM crops or produced by different methods. It is now widely recognised that herbicide dependency must be reduced (Harker et al., 2017; Beckie et al., 2019b).

Viable alternatives include:

- Increased use of agro-ecological methods, for conventional as well as organic farming, including crop rotations;
- Further development and implementation of spot spraying and precision weeding to target and reduce the use of herbicides and/or technologies to limit weed seed production during the grain harvest (Quartz, 2015; Guardian, 2015; Horticulture Week, 2015; Gonzalez-de-Santos et al., 2017; Walsh et al., 2017; Beckie et al., 2019b; Oliver, 2020; Belton, 2021; Peters, 2021).

Even advocates of GM crops now accept that RR crops – the main GM crops that are grown today - are not the future of agriculture. Former UK Life Sciences Minister, George Freeman MP (Minister for Science, Research and Innovation until July 2022) stated: *“The first generation, if you like ‘GM1.0’, was very crude, particularly the original Monsanto monoculture model: “Spray everything that dies apart from the thing we have protected.” I do not think anyone thinks that is a particularly progressive way of doing 21st century agriculture...”* (House of Commons Science and Technology Committee, 2016). Former senior scientists at DuPont and Corteva

Agriscience conclude a recent book chapter, “*Today, glyphosate-based crop systems are still mainstays of weed management, but they cannot keep up with the capacity of weeds to evolve resistance. Growers desperately need new technologies, but no technology with the impact of glyphosate and GR crops is on the horizon. Although the expansion of GR crop traits is possible into new geographic areas and crops such as wheat and sugarcane and could have high value, the Roundup Ready® revolution is over*” (Green & Siehl, 2021).

There are significant opportunity costs associated with investing in ‘next-generation’ HT crops which are tolerant to more herbicides but which will not solve the long-term problem of resistant weeds and will continue to pose risks to human health and the environment. Investing in alternatives means a supporting a paradigm shift towards using less herbicide, not more, to the benefit of farmers, human health and the environment. It is particularly important that RR crops are not pushed into new countries which have so far avoided stepping onto the “transgenic treadmill”.

## **Recommendations**

GeneWatch calls for an end to the cultivation of herbicide-tolerant (HT) GM crops, including RoundUp Ready (RR) crops and ‘next-generation’ GM crops that are tolerant to more than one weedkiller. Protecting the environment and human health should be a priority.

The growing failure of RoundUp Ready crops, due to the spread of glyphosate resistant (GR) weeds, provides an opportunity to phase out the use of RR crops and adopt new methods and technologies. The priority should be to reduce and replace the use of herbicides: not to replace RR crops with other herbicide-tolerant crops, whether or not these are GM crops or produced by different methods.

Governments of countries where RR crops are grown should urgently develop phase-out plans for this technology and publish these for public consultation and debate.

Governments should also end subsidies for maize (corn) to be used as biofuels (corn ethanol), rather than as food.

Governments of countries where RR crops are not currently grown (for example in Europe, most of Africa and Asia, parts of Latin America and New Zealand) should maintain their *de facto* bans on this technology.

In addition:

- Food retailers should require non-GM feed for meat and dairy products, to seek to minimise environmental damage in countries where GM HT crops are grown. At minimum, labelled non-GM-fed meat and dairy products should be available to allow consumers to choose to eat such products.
- In the United States, GM food products should be labelled and food manufacturers should seek to avoid using ingredients from GM HT crops.

## References

- Abarikwu, S. O., Akiri, O. F., Durojaiye, M. A., & Adenike, A. (2015). Combined effects of repeated administration of Bretmont Wipeout (glyphosate) and Ultrazin (atrazine) on testosterone, oxidative stress and sperm quality of Wistar rats. *Toxicology Mechanisms and Methods*, 25(1), 70–80. <http://doi.org/10.3109/15376516.2014.989349>
- Abdul, K. S. M., De Silva, P. M. C. S., Ekanayake, E. M. D. V., Thakshila, W. A. K. G., Gunarathna, S. D., Gunasekara, T. D. K. S. C., Jayasinghe, S. S., Asanthi, H. B., Chandana, E. P. S., Chaminda, G. G. T., Siribaddana, S. H., & Jayasundara, N. (2021). Occupational Paraquat and Glyphosate Exposure May Decline Renal Functions among Rural Farming Communities in Sri Lanka. *International Journal of Environmental Research and Public Health*, 18(6), 3278. <https://doi.org/10.3390/ijerph18063278>
- ACRE. (2019). Advice on an application for deliberate release of a GMO for research and development purposes. Advisory Committee on Releases to the Environment. Notification reference: 18/R8/01. [https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment\\_data/file/800590/gmo-camelina-acre-advice-18r801.pdf](https://assets.publishing.service.gov.uk/government/uploads/system/uploads/attachment_data/file/800590/gmo-camelina-acre-advice-18r801.pdf)
- Acquavella, J. F., Alexander, B. H., Mandel, J. S., Gustin, C., Baker, B., Chapman, P., & Bleeke, M. (2004). Glyphosate biomonitoring for farmers and their families: results from the Farm Family Exposure Study. *Environmental Health Perspectives*, 112(3), 321–326.
- African Centre for Biodiversity. (2021). *Bayer Breathing Life Into Gates' Failed GM Drought Tolerant Maize*. <https://acbio.org.za/wp-content/uploads/2022/04/bayer-breathing-life-gates-failed-gm-drought-tolerant-maizeagrarian-extractivism-continues-unabated.pdf>
- Agostini, L. P., Dettogni, R. S., dos Reis, R. S., Stur, E., dos Santos, E. V. W., Ventrone, D. P., Garcia, F. M., Cardoso, R. C., Graceli, J. B., & Louro, I. D. (2020). Effects of glyphosate exposure on human health: Insights from epidemiological and in vitro studies. *Science of The Total Environment*, 705, 135808. <https://doi.org/10.1016/j.scitotenv.2019.135808>
- Albisser Vögeli, G., Burose, F., Wolf, D., Lips, M. (2011). Wirtschaftlichkeit gentechnisch-veränderter Ackerkulturen in der Schweiz. Mit detaillierter Berücksichtigung möglicher Koexistenz-Kosten. Forschungsanstalt Agroscope Reckenholz-Tänikon ART.
- Alcántara de la Cruz, R., Moraes de Oliveira, G., Carvalho, L., & Silva, M. (2020). *Herbicide Resistance in Brazil: Status, Impacts, and Future Challenges* (pp. 1–25). <https://doi.org/10.5772/intechopen.91236>
- Allison, R. (2015). How glyphosate-resistant weeds will ingest all US crops by 2020. *Farmers Weekly*. <http://www.fwi.co.uk/arable/glyphosate-resistant-weeds-will-infest-all-us-crops-by-2020.htm?cmpid=SOC%7CTwitter%7CFarmersWeekly%7Csf14292641%7Csf14292641>
- Alonso, L. L., Demetrio, P. M., Agustina Etchegoyen, M., & Marino, D. J. (2018). Glyphosate and atrazine in rainfall and soils in agroproductive areas of the pampas region in Argentina. *Science of The Total Environment*, 645, 89–96. <https://doi.org/10.1016/j.scitotenv.2018.07.134>
- Anifandis, G., Amiridis, G., Dafopoulos, K., Daponte, A., Dovolou, E., Gavriil, E., Gorgogietas, V., Kachpani, E., Mamuris, Z., Messini, C. I., Vassiou, K., & Psarra, A.-M. G. (2017). The In Vitro Impact of the Herbicide Roundup on Human Sperm Motility and Sperm Mitochondria. *Toxics*, 6(1), 2. <https://doi.org/10.3390/toxics6010002>
- Anifandis, G., Katsanaki, K., Lagodonti, G., Messini, C., Simopoulou, M., Dafopoulos, K., & Daponte, A. (2018). The Effect of Glyphosate on Human Sperm Motility and Sperm DNA Fragmentation. *International Journal of Environmental Research and Public Health*, 15(6), 1117. <https://doi.org/10.3390/ijerph15061117>
- Arbuckle, T. E., Lin, Z., & Mery, L. S. (2001). An exploratory analysis of the effect of pesticide exposure on the risk of spontaneous abortion in an Ontario farm population. *Environmental Health Perspectives*, 109(8), 851–857.
- Areal, F.J., Riesgo, L., Rodriguez-Cerezo, E. (2013). Economic and agronomic impact of commercialized GM crops: A meta-analysis. *Journal of Agricultural Science* 151:7-33.
- Arregui, M. C., Lenardón, A., Sanchez, D., Maitre, M. I., Scotta, R., & Enrique, S. (2004). Monitoring glyphosate residues in transgenic glyphosate-resistant soybean. *Pest Management Science*, 60(2), 163–166. <http://doi.org/10.1002/ps.775>
- Avigliano, L., Fassiano, A. V., Medesani, D. A., Ríos de Molina, M. C., & Rodríguez, E. M. (2014a). Effects of Glyphosate on Growth Rate, Metabolic Rate and Energy Reserves of Early Juvenile Crayfish, *Cherax quadricarinatus* M. *Bulletin of Environmental Contamination and Toxicology*, 92(6), 631–635. <http://doi.org/10.1007/s00128-014-1240-7>
- Avigliano, L., Alvarez, N., Loughlin, C. M., & Rodríguez, E. M. (2014b). Effects of glyphosate on egg incubation, larvae hatching, and ovarian rematuration in the estuarine crab *Neohelice granulata*. *Environmental Toxicology and Chemistry / SETAC*, 33(8), 1879–1884. <http://doi.org/10.1002/etc.2635>
- Babalola, O. O., Truter, J. C., Archer, E., & van Wyk, J. H. (2021). Exposure Impacts of Environmentally Relevant Concentrations of a Glufosinate Ammonium Herbicide Formulation on Larval Development and Thyroid

- Histology of *Xenopus laevis*. *Archives of Environmental Contamination and Toxicology*, 80(4), 717–725. <https://doi.org/10.1007/s00244-020-00758-3>
- Babich, R., Ulrich, J. C., Ekanayake, E. M. D. V., Massarsky, A., De Silva, P. M. C. S., Manage, P. M., Jackson, B. P., Ferguson, P. L., Di Giulio, R. T., Drummond, I. A., & Jayasundara, N. (2020). Kidney developmental effects of metal-herbicide mixtures: Implications for chronic kidney disease of unknown etiology. *Environment International*, 144, 106019. <https://doi.org/10.1016/j.envint.2020.106019>
- Bach, N. C., Natale, G. S., Somoza, G. M., & Ronco, A. E. (2016). Effect on the growth and development and induction of abnormalities by a glyphosate commercial formulation and its active ingredient during two developmental stages of the South-American Creole frog, *Leptodactylus latrans*. *Environmental Science and Pollution Research International*, 23(23), 23959–23971. <https://doi.org/10.1007/s11356-016-7631-z>
- Barbosa Lima, I., Boechat, I., & Gücker, B. (2021). Glyphosate in Brazil: Use, aquatic contamination, environmental effects, and health hazards. *Caderno de Geografia*, 31, 90–115. <https://doi.org/10.5752/p.2318-2962.2021v31nesp1p90>
- Barnett, J. A., Bandy, M. L., & Gibson, D. L. (2022). Is the Use of Glyphosate in Modern Agriculture Resulting in Increased Neuropsychiatric Conditions Through Modulation of the Gut-brain-microbiome Axis? *Frontiers in Nutrition*, 9. <https://www.frontiersin.org/article/10.3389/fnut.2022.827384>
- Battaglin, W. A., Kolpin, D. W., Scribner, E. A., Kuivila, K. M., & Sandstrom, M. W. (2005). Glyphosate, Other Herbicides, and Transformation Products in Midwestern Streams, 2002. *JAWRA Journal of the American Water Resources Association*, 41(2), 323–332. <http://doi.org/10.1111/j.1752-1688.2005.tb03738.x>
- Battaglin, W. A., Meyer, M. T., Kuivila, K. M., & Dietze, J. E. (2014). Glyphosate and Its Degradation Product AMPA Occur Frequently and Widely in U.S. Soils, Surface Water, Groundwater, and Precipitation. *JAWRA Journal of the American Water Resources Association*, 50(2), 275–290. <http://doi.org/10.1111/jawr.12159>
- Battisti, L., Potrich, M., Sampaio, A. R., de Castilhos Ghisi, N., Costa-Maia, F. M., Abati, R., Dos Reis Martinez, C. B., & Sofia, S. H. (2021). Is glyphosate toxic to bees? A meta-analytical review. *The Science of the Total Environment*, 767, 145397. <https://doi.org/10.1016/j.scitotenv.2021.145397>
- Beckie, H. J., Ashworth, M. B., & Flower, K. C. (2019b). Herbicide Resistance Management: Recent Developments and Trends. *Plants*, 8(6), 161. <https://doi.org/10.3390/plants8060161>
- Bednářová, A., Kropf, M., & Krishnan, N. (2020). The surfactant polyethoxylated tallowamine (POEA) reduces lifespan and inhibits fecundity in *Drosophila melanogaster*- In vivo and in vitro study. *Ecotoxicology and Environmental Safety*, 188, 109883. <https://doi.org/10.1016/j.ecoenv.2019.109883>
- Bellaloui, N., Zablotowicz, R. M., Reddy, K. N., & Abel, C. A. (2008). Nitrogen Metabolism and Seed Composition As Influenced by Glyphosate Application in Glyphosate-Resistant Soybean. *Journal of Agricultural and Food Chemistry*, 56(8), 2765–2772. <http://doi.org/10.1021/jf703615m>
- Belsky, J., & Joshi, N. K. (2018). Assessing Role of Major Drivers in Recent Decline of Monarch Butterfly Population in North America. *Frontiers in Environmental Science*, 0. <https://doi.org/10.3389/fenvs.2018.00086>
- Belton, P. (2021, August 14). 'Ten years ago this was science fiction': The rise of weedkilling robots. *The Guardian*. <https://www.theguardian.com/environment/2021/aug/14/weedkilling-robots-farming-pesticide-use-sustainable>
- Benachour, N., Sipahutar, H., Moslemi, S., Gasnier, C., Travert, C., & Séralini, G. E. (2007). Time- and Dose-Dependent Effects of Roundup on Human Embryonic and Placental Cells. *Archives of Environmental Contamination and Toxicology*, 53(1), 126–133. <http://doi.org/10.1007/s00244-006-0154-8>
- Benachour, N., & Séralini, G.-E. (2009). Glyphosate Formulations Induce Apoptosis and Necrosis in Human Umbilical, Embryonic, and Placental Cells. *Chemical Research in Toxicology*, 22(1), 97–105. <http://doi.org/10.1021/tx800218n>
- Benbrook, C. M. (2012a). Impacts of genetically engineered crops on pesticide use in the U.S. -- the first sixteen years. *Environmental Sciences Europe*, 24(24), 2190–4715. <http://doi.org/10.1186/2190-4715-24-24>
- Benbrook, C. M. (2016). Trends in glyphosate herbicide use in the United States and globally. *Environmental Sciences Europe*, 28(1). <http://doi.org/10.1186/s12302-016-0070-0>
- Benbrook, C. M. (2018). Why Regulators Lost Track and Control of Pesticide Risks: Lessons From the Case of Glyphosate-Based Herbicides and Genetically Engineered-Crop Technology. *Current Environmental Health Reports*, 5(3), 387–395. <https://doi.org/10.1007/s40572-018-0207-y>
- Benítez-Leite, S., Macchi, M. L., & Acosta, M. (2007). Malformaciones congénitas asociadas a agrotóxicos. *Pediatría (Asunción)*, 34(2), 111–121.
- Benoit, L., Hedges, B., Schryver, M. G., Soltani, N., Hooker, D. C., Robinson, D. E., Laforest, M., Soufiane, B., Tranel, P. J., Giacomini, D., & Sikkema, P. H. (2020). The first record of protoporphyrinogen oxidase and four-way herbicide resistance in eastern Canada. *Canadian Journal of Plant Science*, 100(3), 327–331. <https://doi.org/10.1139/cjps-2018-0326>
- Bento, C. P. M., Yang, X., Gort, G., Xue, S., van Dam, R., Zomer, P., Mol, H. G. J., Ritsema, C. J., & Geissen, V. (2016). Persistence of glyphosate and aminomethylphosphonic acid in loess soil under different

- combinations of temperature, soil moisture and light/darkness. *Science of The Total Environment*, 572, 301–311. <https://doi.org/10.1016/j.scitotenv.2016.07.215>
- Berger, M., & Ortega, F. (2010). Populations exposed to agrotoxics: citizens' self-organization in the defense of life and health, Córdoba City, Argentina. *Physis: Revista de Saúde Coletiva*, 20(1), 119–143. <http://doi.org/10.1590/S0103-73312010000100008>
- Berman, M. C., Llamas, M. E., Minotti, P., Fermani, P., Quiroga, M. V., Ferraro, M. A., Metz, S., & Zagarese, H. E. (2020). Field evidence supports former experimental claims on the stimulatory effect of glyphosate on picocyanobacteria communities. *Science of The Total Environment*, 701, 134601. <https://doi.org/10.1016/j.scitotenv.2019.134601>
- Binimelis, R. (2008). Coexistence of Plants and Coexistence of Farmers: Is an Individual Choice Possible? *Journal of Agricultural and Environmental Ethics*, 21(5), 437–457. <http://doi.org/10.1007/s10806-008-9099-4>
- Binimelis, R., Pengue, W., & Monterroso, I. (2009). “Transgenic treadmill”: responses to the emergence and spread of glyphosate-resistant johnsongrass in Argentina. *Geoforum*, 40(4), 623–633. <http://doi.org/10.1016/j.geoforum.2009.03.009>
- Bohm, G.M.B., Rombaldi, C. V., Genovese, M. I., Castilhos, D., Alves, R., ... Rumjanek, N. G. (2014). Glyphosate Effects on Yield, Nitrogen Fixation, and Seed Quality in Glyphosate-Resistant Soybean. *Crop Science*, 54(4), 1737–1743. <http://doi.org/10.2135/cropsci2013.07.0470>
- Bøhn, T., Cuhra, M., Traavik, T., Sanden, M., Fagan, J., & Primicerio, R. (2014). Compositional differences in soybeans on the market: Glyphosate accumulates in Roundup Ready GM soybeans. *Food Chemistry*, 153, 207–215. <http://doi.org/10.1016/j.foodchem.2013.12.054>
- Bøhn, T., & Millstone, E. (2019). The Introduction of Thousands of Tonnes of Glyphosate in the food Chain—An Evaluation of Glyphosate Tolerant Soybeans. *Foods*, 8(12), 669. <https://doi.org/10.3390/foods8120669>
- Bonansea, R. I., Filippi, I., Wunderlin, D. A., Marino, D. J. G., & Amé, M. V. (2017). The Fate of Glyphosate and AMPA in a Freshwater Endorheic Basin: An Ecotoxicological Risk Assessment. *Toxics*, 6(1), 3. <https://doi.org/10.3390/toxics6010003>
- Bott, S., Tesfamariam, T., Candan, H., Cakmak, I., Römheld, V., & Neumann, G. (2008). Glyphosate-induced impairment of plant growth and micronutrient status in glyphosate-resistant soybean (*Glycine max* L.). *Plant and Soil*, 312(1-2), 185–194. <http://doi.org/10.1007/s11104-008-9760-8>
- Botten, N., Wood, L. J., & Werner, J. R. (2021). Glyphosate remains in forest plant tissues for a decade or more. *Forest Ecology and Management*, 493, 119259. <https://doi.org/10.1016/j.foreco.2021.119259>
- Brausch, J. M., Beall, B., & Smith, P. N. (2007). Acute and Sub-Lethal Toxicity of Three POEA Surfactant Formulations to *Daphnia magna*. *Bulletin of Environmental Contamination and Toxicology*, 78(6), 510–514. <http://doi.org/10.1007/s00128-007-9091-0>
- Brausch, J. M., & Smith, P. N. (2007). Toxicity of Three Polyethoxylated Tallowamine Surfactant Formulations to Laboratory and Field Collected Fairy Shrimp, *Thamnocephalus platyurus*. *Archives of Environmental Contamination and Toxicology*, 52(2), 217–221. <http://doi.org/10.1007/s00244-006-0151-y>
- Brovini, E. M., de Deus, B. C. T., Vilas-Boas, J. A., Quadra, G. R., Carvalho, L., Mendonça, R. F., Pereira, R. de O., & Cardoso, S. J. (2021a). Three-best-seller pesticides in Brazil: Freshwater concentrations and potential environmental risks. *Science of The Total Environment*, 771, 144754. <https://doi.org/10.1016/j.scitotenv.2020.144754>
- Brovini, E. M., Cardoso, S. J., Quadra, G. R., Vilas-Boas, J. A., Paranaíba, J. R., Pereira, R. de O., & Mendonça, R. F. (2021b). Glyphosate concentrations in global freshwaters: Are aquatic organisms at risk? *Environmental Science and Pollution Research*, 28(43), 60635–60648. <https://doi.org/10.1007/s11356-021-14609-8>
- Brower, L. P., Taylor, O. R., Williams, E. H., Slayback, D. A., Zubieta, R. R., & Ramírez, M. I. (2012). Decline of monarch butterflies overwintering in Mexico: is the migratory phenomenon at risk?: Decline of monarch butterflies in Mexico.
- Brunharo, C. A. C. G., Gast, R., Kumar, V., Mallory-Smith, C. A., Tidemann, B. D., & Beckie, H. J. (2022). Western United States and Canada perspective: Are herbicide-resistant crops the solution to herbicide-resistant weeds? *Weed Science*, 70(3), 272–286. <https://doi.org/10.1017/wsc.2022.6>
- Bruns, H. A. (2014). Stacked-Gene Hybrids Were Not Found to Be Superior to Glyphosate-Resistant or Non-GMO Corn Hybrids. *Crop Management*, 13(1), 0. <http://doi.org/10.2134/CM-2013-0012-RS>
- Bunge, J. (2015). Fields of Gold: GMO-Free Crops Prove Lucrative for Farmers. *The Wall Street Journal*. <http://www.wsj.com/articles/fields-of-gold-gmo-free-crops-prove-lucrative-for-farmers-1422909700>
- Burchfield, E. K., Schumacher, B. L., Spangler, K., & Rissing, A. (2022). The State of US Farm Operator Livelihoods. *Frontiers in Sustainable Food Systems*, 5. <https://www.frontiersin.org/article/10.3389/fsufs.2021.795901>
- Burgeff, C., Huerta, E., Acevedo, F., & Sarukhán, J. (2014). How Much Can GMO and Non-GMO Cultivars Coexist in a Megadiverse Country? Retrieved December 15, 2014, from <http://www.agbioforum.org/v17n1/v17n1a10-burgeff.htm>

- Burke (2003). GM crops. Effects on Farmland Wildlife. Farm Scale Evaluations. <http://webarchive.nationalarchives.gov.uk/20080306073937/http://www.defra.gov.uk/environment/gm/fse/reports/fse-summary-03.pdf>
- Burke (2005). Managing GM crops with herbicides. Effects on farmland wildlife. Farm Scale Evaluations. <http://webarchive.nationalarchives.gov.uk/20080306073937/http://www.defra.gov.uk/environment/gm/fse/reports/fse-summary-05.pdf>
- Cai, W., Ji, Y., Song, X., Guo, H., Han, L., Zhang, F., Liu, X., Zhang, H., Zhu, B., & Xu, M. (2017). Effects of glyphosate exposure on sperm concentration in rodents: A systematic review and meta-analysis. *Environmental Toxicology and Pharmacology*, 55, 148–155. <https://doi.org/10.1016/j.etap.2017.07.015>
- Camiccia, M., Candioto, L. Z. P., Gaboardi, S. C., Panis, C., & Kottwitz, L. B. M. (2022). Determination of glyphosate in breast milk of lactating women in a rural area from Paraná state, Brazil. *Brazilian Journal of Medical and Biological Research*, 55. <https://doi.org/10.1590/1414-431X2022e12194>
- Carretta, L., Masin, R., & Zanin, G. (2022). Review of studies analysing glyphosate and aminomethylphosphonic acid (AMPA) occurrence in groundwater. *Environmental Reviews*, 30(1), 88–109. <https://doi.org/10.1139/er-2020-0106>
- Cassidy, E. S., West, P. C., Gerber, J. S., & Foley, J. A. (2013). Redefining agricultural yields: from tonnes to people nourished per hectare. *Environmental Research Letters*, 8(3), 034015. <http://doi.org/10.1088/1748-9326/8/3/034015>
- Cassidy, E. (2015a). How Corn Ethanol Is Worse For Climate Change Than The Keystone Pipeline. Environmental Working Group. <http://www.ewg.org/agmag/2015/05/how-corn-ethanol-worse-climate-change-keystone-pipeline>
- Cassidy, E. (2015b). Feeding the World Without GMOs. Environmental Working Group. <http://www.ewg.org/research/feeding-world-without-gmos>
- Caulcutt, C. (2009). 'Superweed' explosion threatens Monsanto heartlands. France 24. Published April 19, 2009. <http://www.france24.com/en/20090419-'superweed'-explosion-threatens-monsanto-heartlands->
- Cedergreen, N., & Streibig, J. C. (2005). The toxicity of herbicides to non-target aquatic plants and algae: assessment of predictive factors and hazard. *Pest Management Science*, 61(12), 1152–1160. <http://doi.org/10.1002/ps.1117>
- Cederlund, H. (2017). Effects of spray drift of glyphosate on nontarget terrestrial plants—A critical review. *Environmental Toxicology and Chemistry*, 36(11), 2879–2886. <https://doi.org/10.1002/etc.3925>
- Cerdeira, A. L., Gazziero, D. L. P., Duke, S. O., & Matallo, M. B. (2011). Agricultural Impacts of Glyphosate-Resistant Soybean Cultivation in South America. *Journal of Agricultural and Food Chemistry*, 59(11), 5799–5807. <http://doi.org/10.1021/jf102652y>
- Chang, F., Simcik, M. F., & Capel, P. D. (2011). Occurrence and fate of the herbicide glyphosate and its degradate aminomethylphosphonic acid in the atmosphere. *Environmental Toxicology and Chemistry*, 30(3), 548–555. <http://doi.org/10.1002/etc.431>
- Chávez-Ortiz, P., Tapia-Torres, Y., Larsen, J., & García-Oliva, F. (2022). Glyphosate-based herbicides alter soil carbon and phosphorus dynamics and microbial activity. *Applied Soil Ecology*, 169, 104256. <https://doi.org/10.1016/j.apsoil.2021.104256>
- Chen, J., Jin, M., Qiu, Z.-G., Guo, C., Chen, Z.-L., Shen, Z.-Q., Wang, X.-W., Li, J.-W. (2012). A survey of drug resistance bla genes originating from synthetic plasmid vectors in six Chinese rivers. *Environmental Science & Technology*, 46(24), 13448–13454. <http://doi.org/10.1021/es302760s>
- Chłopecka, M., Mendel, M., Dziekan, N., Karlik, W. (2017). The effect of glyphosate-based herbicide Roundup and its co-formulant, POEA, on the motoric activity of rat intestine – In vitro study. *Environmental Toxicology and Pharmacology* 49, 156–162. doi:10.1016/j.etap.2016.12.010
- Cipriano, J., Carrasco, J. F., Arbós, M. (2006). La imposible coexistencia: Siete años de transgénicos contaminan el maíz ecológico y el convencional: una aproximación a partir de los casos de Cataluña y Aragón. Asamblea PAGESA de Catalunya, Greenpeace, Plataforma Transgènics Fora.
- Clair, É., Mesnage, R., Travert, C., & Séralini, G.-É. (2012). A glyphosate-based herbicide induces necrosis and apoptosis in mature rat testicular cells in vitro, and testosterone decrease at lower levels. *Toxicology in Vitro*, 26(2), 269–279. <http://doi.org/10.1016/j.tiv.2011.12.009>
- Clapp, J. (2021). The problem with growing corporate concentration and power in the global food system. *Nature Food*, 2(6), 404–408. <https://doi.org/10.1038/s43016-021-00297-7>
- Clasen, B., Murussi, C., & Storck, T. (2019). Pesticide Contamination in Southern Brazil. In L. M. Gómez-Oliván (Ed.), *Pollution of Water Bodies in Latin America: Impact of Contaminants on Species of Ecological Interest* (pp. 43–54). Springer International Publishing. [https://doi.org/10.1007/978-3-030-27296-8\\_3](https://doi.org/10.1007/978-3-030-27296-8_3)
- Comont, D., Lowe, C., Hull, R., Crook, L., Hicks, H. L., Onkokesung, N., Beffa, R., Childs, D. Z., Edwards, R., Freckleton, R. P., & Neve, P. (2020). Evolution of generalist resistance to herbicide mixtures reveals a trade-off in resistance management. *Nature Communications*, 11, 3086. <https://doi.org/10.1038/s41467-020-16896-0>

- Cox, C., & Sorgan, M. (2006). Unidentified Inert Ingredients in Pesticides: Implications for Human and Environmental Health. *Environmental Health Perspectives*, 114(12), 1803–1806. <http://doi.org/10.1289/ehp.9374>
- Cristofaro, C. da S., Branco, C. W. C., Rocha, M. I. de A., & Portugal, S. da G. M. (2021). Assessing glyphosate concentrations in six reservoirs of Paraíba do Sul and Guandu River Basins in southeast Brazil. *Revista Ambiente & Água*, 16. <https://doi.org/10.4136/ambi-agua.2615>
- Cuhra, M., Traavik, T., & Bøhn, T. (2013). Clone- and age-dependent toxicity of a glyphosate commercial formulation and its active ingredient in *Daphnia magna*. *Ecotoxicology*, 22(2), 251–262. <http://doi.org/10.1007/s10646-012-1021-1>
- Cuhra, M., Traavik, T., & Bøhn, T. (2014). Life cycle fitness differences in *Daphnia magna* fed Roundup-Ready soybean or conventional soybean or organic soybean. *Aquaculture Nutrition*, 21(5), 702–713. <http://doi.org/10.1111/anu.12199>
- Cuhra, M., Traavik, T., Dando, M., Primicerio, R., Holderbaum, D. F., & Bøhn, T. (2015). Glyphosate-Residues in Roundup-Ready Soybean Impair *Daphnia magna* Life-Cycle. *Journal of Agricultural Chemistry and Environment*, 04(01), 24–36. <http://doi.org/10.4236/jacen.2015.41003>
- Dabney, B. L., & Patiño, R. (2018). Low-dose stimulation of growth of the harmful alga, *Prymnesium parvum*, by glyphosate and glyphosate-based herbicides. *Harmful Algae*, 80, 130–139. <https://doi.org/10.1016/j.hal.2018.11.004>
- da Costa, N. B., Hébert, M.-P., Fugère, V., Terrat, Y., Fussmann, G. F., Gonzalez, A., & Shapiro, B. J. (2021). A glyphosate-based herbicide cross-selects for antibiotic resistance genes in bacterioplankton communities (p. 2021.12.13.472531). bioRxiv. <https://doi.org/10.1101/2021.12.13.472531>
- Daisley, B. A., Chernyshova, A. M., Thompson, G. J., & Allen-Vercoe, E. (2022). Deteriorating microbiomes in agriculture—The unintended effects of pesticides on microbial life. *Microbiome Research Reports*, 1(1), 6. <https://doi.org/10.20517/mrr.2021.08>
- da Silva, K. A., Nicola, V. B., Dudas, R. T., Demetrio, W. C., Maia, L. dos S., Cunha, L., Bartz, M. L. C., Brown, G. G., Pasini, A., Kille, P., Ferreira, N. G. C., & de Oliveira, C. M. R. (2021). Pesticides in a case study on no-tillage farming systems and surrounding forest patches in Brazil. *Scientific Reports*, 11(1), 9839. <https://doi.org/10.1038/s41598-021-88779-3>
- da Silva, A. P., Morais, E. R., Oliveira, E. C., & Ghisi, N. de C. (2022). Does exposure to environmental 2,4-dichlorophenoxyacetic acid concentrations increase mortality rate in animals? A meta-analytic review. *Environmental Pollution*, 303, 119179. <https://doi.org/10.1016/j.envpol.2022.119179>
- Davies, S. (2020, February 19). *Following \$265M peach farm verdict, dicamba litigation may expand*. <https://www.agripulse.com/articles/13197-following-265m-peach-farm-verdict-dicamba-litigation-may-expand?v=preview>
- de Brito Rodrigues, L., Gonçalves Costa, G., Lundgren Thá, E., da Silva, L. R., de Oliveira, R., Morais Leme, D., Cestari, M. M., Koppe Grisolia, C., Campos Valadares, M., & de Oliveira, G. A. R. (2019). Impact of the glyphosate-based commercial herbicide, its components and its metabolite AMPA on non-target aquatic organisms. *Mutation Research/Genetic Toxicology and Environmental Mutagenesis*, 842, 94–101. <https://doi.org/10.1016/j.mrgentox.2019.05.002>
- Dedeke, G. A., Owagboriaye, F. O., Ademolu, K. O., Olujimi, O. O., & Aladesida, A. A. (2018). Comparative Assessment on Mechanism Underlying Renal Toxicity of Commercial Formulation of Roundup Herbicide and Glyphosate Alone in Male Albino Rat. *International Journal of Toxicology*, 37(4), 285–295. <https://doi.org/10.1177/1091581818779553>
- Defarge, N., Spiroux de Vendômois, J., & Séralini, G. E. (2018). Toxicity of formulants and heavy metals in glyphosate-based herbicides and other pesticides. *Toxicology Reports*, 5, 156–163. <https://doi.org/10.1016/j.toxrep.2017.12.025>
- de Groot, G. S., Aizen, M. A., Sáez, A., & Morales, C. L. (2021). Large-scale monoculture reduces honey yield: The case of soybean expansion in Argentina. *Agriculture, Ecosystems & Environment*, 306, 107203. <https://doi.org/10.1016/j.agee.2020.107203>
- De la Fuente, E. B., Suárez, S. A., & Ghersa, C. M. (2006). Soybean weed community composition and richness between 1995 and 2003 in the Rolling Pampas (Argentina). *Agriculture, Ecosystems & Environment*, 115(1–4), 229–236. <http://doi.org/10.1016/j.agee.2006.01.009>
- De La Fuente, E. B., Perelman, S., & Ghersa, C. M. (2010). Weed and arthropod communities in soybean as related to crop productivity and land use in the Rolling Pampa, Argentina. *Weed Research*, 50(6), 561–571. <http://doi.org/10.1111/j.1365-3180.2010.00811.x>
- de María, M., Silva-Sanchez, C., Kroll, K. J., Walsh, M. T., Nouri, M.-Z., Hunter, M. E., Ross, M., Clauss, T. M., & Denslow, N. D. (2021). Chronic exposure to glyphosate in Florida manatee. *Environment International*, 152, 106493. <https://doi.org/10.1016/j.envint.2021.106493>
- de Maria, M., Kroll, K. J., Yu, F., Nouri, M.-Z., Silva-Sanchez, C., Perez, J. G., Moraga Amador, D. A., Zhang, Y., Walsh, M. T., & Denslow, N. D. (2022). Endocrine, immune and renal toxicity in male largemouth bass after chronic exposure to glyphosate and Rodeo®. *Aquatic Toxicology*, 246, 106142. <https://doi.org/10.1016/j.aquatox.2022.106142>

- de Souza, A. P. F., Rodrigues, N. R., & Reyes, F. G. R. (2021). Glyphosate and aminomethylphosphonic acid (AMPA) residues in Brazilian honey. *Food Additives & Contaminants: Part B*, 14(1), 40–47. <https://doi.org/10.1080/19393210.2020.1855676>
- Desquilbet, M., Bullock, D. S., & D'Arcangelo, F. M. (2019). A discussion of the market and policy failures associated with the adoption of herbicide-tolerant crops. *International Journal of Agricultural Sustainability*, 17(5), 326–337. <https://doi.org/10.1080/14735903.2019.1655191>
- Dias, M., Rocha, R., & Soares, R. R. (2020). *Down the River: Glyphosate Use in Agriculture and Birth Outcomes of Surrounding Populations*. Latin American And The Caribbean Economic Association. [http://vox.lacea.org/files/Working\\_Papers/lacea\\_wps\\_0024\\_dias\\_rocha\\_soares.pdf](http://vox.lacea.org/files/Working_Papers/lacea_wps_0024_dias_rocha_soares.pdf)
- Dick, R. E., & Quinn, J. P. (1995). Glyphosate-degrading isolates from environmental samples: occurrence and pathways of degradation. *Applied Microbiology and Biotechnology*, 43(3), 545–550. <http://doi.org/10.1007/BF00218464>
- Doering, C. (2015). Farmers turn to GMO-free crops to boost income. Press Citizen. <http://www.press-citizen.com/story/money/agriculture/2015/04/18/non-gmo-farming/25951693/>
- Domínguez, A., Brown, G. G., Sautter, K. D., Ribas de Oliveira, C. M., de Vasconcelos, E. C., Niva, C. C., Bartz, M. L., Bedano, J. C. (2016). Toxicity of AMPA to the earthworm *Eisenia andrei* Bouché, 1972 in tropical artificial soil. *Scientific Reports*, 6, 19731. <http://doi.org/10.1038/srep19731>
- Dreoni, I., Matthews, Z., & Schaafsma, M. (2022). The impacts of soy production on multi-dimensional well-being and ecosystem services: A systematic review. *Journal of Cleaner Production*, 335, 130182. <https://doi.org/10.1016/j.jclepro.2021.130182>
- Duke, S. O., Powles, S. B. (2008). Glyphosate: a once-in-a-century herbicide. *Pest Management Science* 64: 319-325.
- Duke, S. O. (2018). The history and current status of glyphosate. *Pest Management Science*, 74(5), 1027–1034. <https://doi.org/10.1002/ps.4652>
- Duke, S. O., Powles, S. B., & Sammons, R. D. (2018). Glyphosate – How it Became a Once in a Hundred Year Herbicide and Its Future. *Outlooks on Pest Management*, 29(6), 247–251. [https://doi.org/10.1564/v29\\_dec\\_03](https://doi.org/10.1564/v29_dec_03)
- EFSA. (2017, December 22). Request for an EFSA peer review (EFSA Conclusion) on the active substance glufosinate according to Article 13 of Regulation (EU) No 844/2012. Register of Questions. <http://registerofquestions.efsa.europa.eu/roqFrontend/questionLoader?question=EFSA-Q-2016-00408>
- Eker, S., Ozturk, L., Yazici, A., Erenoglu, B., Romheld, V., & Cakmak, I. (2006). Foliar-Applied Glyphosate Substantially Reduced Uptake and Transport of Iron and Manganese in Sunflower (*Helianthus annuus* L.) Plants. *Journal of Agricultural and Food Chemistry*, 54(26), 10019–10025. <http://doi.org/10.1021/jf0625196>
- Elgert, L. (2016). “More soy on fewer farms” in Paraguay: challenging neoliberal agriculture’s claims to sustainability. *The Journal of Peasant Studies*, 43(2), 537–561. <http://doi.org/10.1080/03066150.2015.1076395>
- Elmore, B. (2022, January 26). The herbicide dicamba was supposed to solve farmers’ weed problems – instead, it’s making farming harder for many of them. *The Conversation*. <http://theconversation.com/the-herbicide-dicamba-was-supposed-to-solve-farmers-weed-problems-instead-its-making-farming-harder-for-many-of-them-174181>
- Eloy, L., Aubertin, C., Toni, F., Lúcio, S. L. B., & Bosgiraud, M. (2016). On the margins of soy farms: Traditional populations and selective environmental policies in the Brazilian Cerrado. *The Journal of Peasant Studies*, 43(2), 494–516. <https://doi.org/10.1080/03066150.2015.1013099>
- EU/FAO/WFP. (2022). Global Report on Food Crises: Acute food insecurity hits new highs. <https://www.fao.org/newsroom/detail/global-report-on-food-crises-acute-food-insecurity-hits-new-highs/en>
- European Commission. (n.d.). *EU Pesticides database—Glufosinate*. European Commission Website. [https://ec.europa.eu/food/plant/pesticides/eu-pesticides-database/active-substances/?event=as.details&as\\_id=79](https://ec.europa.eu/food/plant/pesticides/eu-pesticides-database/active-substances/?event=as.details&as_id=79)
- Evans, J. A., Tranel, P. J., Hager, A. G., Schutte, B., Wu, C., Chatham, L. A., & Davis, A. S. (2016). Managing the evolution of herbicide resistance. *Pest Management Science*, 72(1), 74–80. <http://doi.org/10.1002/ps.4009>
- Everett, K. D. E., & Dickerson, H. W. (2003). *Ichthyophthirius multifiliis* and *tetrahymena thermophila* tolerate glyphosate but not a commercial herbicidal formulation. *Bulletin of Environmental Contamination and Toxicology*, 70(4), 0731–0738. <http://doi.org/10.1007/s00128-003-0044-y>
- Fallon, S. (2014). Monarch butterfly population hits a new low. Switchboard. Natural Resource Defense Council Staff Blog. <https://www.nrdc.org/experts/sylvia-fallon/monarch-butterfly-population-hits-new-low>
- FAO (2011). Global food losses and food waste – Extent, causes and prevention. Rome. [https://knowledge4policy.ec.europa.eu/publication/global-food-losses-food-waste-extent-causes-prevention\\_en](https://knowledge4policy.ec.europa.eu/publication/global-food-losses-food-waste-extent-causes-prevention_en)
- Fargione, J., Hill, J., Tilman, D., Polasky, S., & Hawthorne, P. (2008). Land Clearing and the Biofuel Carbon Debt. *Science*, 319(5867), 1235–1238. <http://doi.org/10.1126/science.1152747>

- Fernandes, G., Aparicio, V. C., Bastos, M. C., De Gerónimo, E., Labanowski, J., Prestes, O. D., Zanella, R., & dos Santos, D. R. (2019). Indiscriminate use of glyphosate impregnates river epilithic biofilms in southern Brazil. *Science of The Total Environment*, 651, 1377–1387. <https://doi.org/10.1016/j.scitotenv.2018.09.292>
- Fernandes, G. B., Silva, A. C. de L., Maronhas, M. E. S., Santos, A. da S. dos, & Lima, P. H. C. (2022). Transgene Flow: Challenges to the On-Farm Conservation of Maize Landraces in the Brazilian Semi-Arid Region. *Plants*, 11(5), 603. <https://doi.org/10.3390/plants11050603>
- Fernandez-Cornejo, J., Wechsler, S., Livingston, M., Mitchel, L. (2014). Genetically Engineered Crops in the United States. ERR-162 USDA, Economic Research Service
- Ferreira, C., Duarte, S. C., Costa, E., Pereira, A. M. P. T., Silva, L. J. G., Almeida, A., Lino, C., & Pena, A. (2021). Urine biomonitoring of glyphosate in children: Exposure and risk assessment. *Environmental Research*, 198, 111294. <https://doi.org/10.1016/j.envres.2021.111294>
- Filomeno, F. (2013). How Argentine Farmers Overpowered Monsanto: The Mobilization of Knowledge-users and Intellectual Property Regimes. *Journal of Politics in Latin America*, 5(3), 35–71.
- Firbank, L. G., Heard, M. S., Woiwod, I. P., Hawes, C., Haughton, A. J., Champion, G. T., Scott, R. J., Hill, M. O., Dewar, A. M., Squire, G. R., May, M. J., Brooks, D. R., Bohan, D. A., Daniels, R. E., Osborne, J. L., Roy, D. B., Black, H. I. J., Rothery, P., Perry, J. N. (2003). An introduction to the Farm-Scale Evaluations of genetically modified herbicide-tolerant crops. *Journal of Applied Ecology*, 40(1), 2–16. <http://doi.org/10.1046/j.1365-2664.2003.00787.x>
- Flockhart, D. T. T., Pichancourt, J.-B., Norris, D. R., & Martin, T. G. (2014). Unravelling the annual cycle in a migratory animal: breeding-season habitat loss drives population declines of monarch butterflies. *Journal of Animal Ecology*, n/a–n/a. <http://doi.org/10.1111/1365-2656.12253>
- Fortune Business Insights. (2022, February). Non-GMO Food Market Size, Trends & Growth | Forecast [2028]. <https://www.fortunebusinessinsights.com/non-gmo-food-market-106359>
- Fraanje, W., & Garnett, T. (2020). *Soy: Food, feed, and land use change*. Food Climate Research Network. <https://www.tabledebates.org/building-blocks/soy-food-feed-and-land-use-change>
- Freisthler, M. S., Robbins, C. R., Benbrook, C. M., Young, H. A., Haas, D. M., Winchester, P. D., & Perry, M. J. (2022). Association between increasing agricultural use of 2,4-D and population biomarkers of exposure: Findings from the National Health and Nutrition Examination Survey, 2001–2014. *Environmental Health*, 21(1), 23. <https://doi.org/10.1186/s12940-021-00815-x>
- Freitas-Silva, L. de, Araújo, H. H., Meireles, C. S., & Silva, L. C. da. (2021). Plant exposure to glyphosate-based herbicides and how this might alter plant physiological and structural processes. *Botany*, 1–8. <https://doi.org/10.1139/cjb-2021-0033>
- Fuchs, B., Saikkonen, K., & Helander, M. (2021). Glyphosate-Modulated Biosynthesis Driving Plant Defense and Species Interactions. *Trends in Plant Science*, 26(4), 312–323. <https://doi.org/10.1016/j.tplants.2020.11.004>
- Furst, I. (1999). Swiss Soiled Seed Prompts Tolerance Question. *Nature Biotechnology*, 17, 629.
- Gabriel, A. & Menrad, K. (2015). Cost of Coexistence of GM and Non-GM Products in the Food Supply Chains of Rapeseed Oil and Maize Starch in Germany. *Agribusiness*, 31(4), 472-490. DOI: 10.1002/agr.21415
- Gaitán-Cremaschi, D., Kamali, F. P., van Evert, F. K., Meuwissen, M. P. M., & Oude Lansink, A. G. J. M. (2015). Benchmarking the sustainability performance of the Brazilian non-GM and GM soybean meal chains: An indicator-based approach. *Food Policy*, 55, 22–32. <http://doi.org/10.1016/j.foodpol.2015.05.006>
- Garrett, R. D., Rueda, X., & Lambin, E. F. (2013). Globalization's unexpected impact on soybean production in South America: linkages between preferences for non-genetically modified crops, eco-certifications, and land use. *Environmental Research Letters*, 8(4), 44055. <http://doi.org/10.1088/1748-9326/8/4/044055>
- Garrett, R. D., & Rausch, L. L. (2016). Green for gold: social and ecological tradeoffs influencing the sustainability of the Brazilian soy industry. *The Journal of Peasant Studies*, 43(2), 461–493. <http://doi.org/10.1080/03066150.2015.1010077>
- Garry, V. F., Harkins, M. E., Erickson, L. L., Long-Simpson, L. K., Holland, S. E., & Burroughs, B. L. (2002). Birth defects, season of conception, and sex of children born to pesticide applicators living in the Red River Valley of Minnesota, USA. *Environmental Health Perspectives*, 110(Suppl 3), 441–449.
- Gasnier, C., Dumont, C., Benachour, N., Clair, E., Chagnon, M.-C., & Séralini, G.-E. (2009). Glyphosate-based herbicides are toxic and endocrine disruptors in human cell lines. *Toxicology*, 262(3), 184–191. <http://doi.org/10.1016/j.tox.2009.06.006>
- Gaupp-Berghausen, M., Hofer, M., Rewald, B., & Zaller, J. G. (2015). Glyphosate-based herbicides reduce the activity and reproduction of earthworms and lead to increased soil nutrient concentrations. *Scientific Reports*, 5, 12886. <http://doi.org/10.1038/srep12886>
- Gazziero, D. L. P. (2015). Mixtures of pesticides in tank, in Brazilian farms. *Planta Daninha*, 33(1), 83–92. <https://doi.org/10.1590/S0100-83582015000100010>
- Gibbons, D. W., Bohan, D. A., Rothery, P., Stuart, R. C., Haughton, A. J., Scott, R. J., Wilson, J. D., Perry, J. N., Clark, S. J., Dawson, R. J. G., Firbank, L. G. (2006). Weed seed resources for birds in fields with contrasting

- conventional and genetically modified herbicide-tolerant crops. *Proceedings of the Royal Society B: Biological Sciences*, 273(1596), 1921–1928. <http://doi.org/10.1098/rspb.2006.3522>
- Gill, J. P. K., Sethi, N., Mohan, A., Datta, S., & Girdhar, M. (2018). Glyphosate toxicity for animals. *Environmental Chemistry Letters*, 16(2), 401–426. <https://doi.org/10.1007/s10311-017-0689-0>
- Gillam, C. (2020b, February 26). *Dicamba litigation against Bayer, BASF poised to explode, lawyers say*. U.S. Right to Know. <https://usrtk.org/monsanto-roundup-trial-tacker/dicamba-litigation-against-bayer-basf-poised-to-explode-lawyers-say/>
- Gillezeau, C., Lieberman-Cribbin, W., & Taioli, E. (2020). Update on human exposure to glyphosate, with a complete review of exposure in children. *Environmental Health*, 19(1), 115. <https://doi.org/10.1186/s12940-020-00673-z>
- Goldfarb, L., & van der Haar, G. (2016). The moving frontiers of genetically modified soy production: shifts in land control in the Argentinian Chaco. *The Journal of Peasant Studies*, 43(2), 562–582. <http://doi.org/10.1080/03066150.2015.1041107>
- Gonçalves, B. B., Giaquinto, P. C., Silva, D. dos S., Neto, C. de M. e S., Lima, A. A. de, Darosci, A. A. B., Portinho, J. L., Carvalho, W. F., & Rocha, T. L. (2019). Ecotoxicology of Glyphosate-Based Herbicides on Aquatic Environment. In *Biochemical Toxicology—Heavy Metals and Nanomaterials*. IntechOpen. <https://doi.org/10.5772/intechopen.85157>
- Gonzalez-de-Santos, P., Ribeiro, A., Fernandez-Quintanilla, C., Lopez-Granados, F., Brandstoeffer, M., Tomic, S., Pedrazzi, S., Peruzzi, A., Pajares, G., Kaplanis, G., Perez-Ruiz, M., Valero, C., del Cerro, J., Vieri, M., Rabatel, G., & Debilde, B. (2017). Fleets of robots for environmentally-safe pest control in agriculture. *Precision Agriculture*, 18(4), 574–614. <https://doi.org/10.1007/s11119-016-9476-3>
- Gosavi, G., Ren, B., Li, X., Zhou, X., Spetz, C., & Zhou, H. (2022). A New Era in Herbicide-Tolerant Crops Development by Targeted Genome Editing. *ACS Agricultural Science & Technology*. <https://doi.org/10.1021/acsagscitech.1c00254>
- Grand View Research. (2019, July). Non-GMO Food Market Size Worth \$2.76 Billion By 2025 | CAGR 16.5%. <https://www.grandviewresearch.com/press-release/global-non-gmo-food-market>
- Grau, D., Grau, N., Gascuel, Q., Paroissin, C., Stratonovitch, C., Lairon, D., Devault, D. A., & Di Cristofaro, J. (2022). Quantifiable urine glyphosate levels detected in 99% of the French population, with higher values in men, in younger people, and in farmers. *Environmental Science and Pollution Research*, 29(22), 32882–32893. <https://doi.org/10.1007/s11356-021-18110-0>
- Green, J. M., & Siehl, D. L. (2021). History and Outlook for Glyphosate-Resistant Crops. In J. B. Knaak (Ed.), *Reviews of Environmental Contamination and Toxicology Volume 255: Glyphosate* (pp. 67–91). Springer International Publishing. [https://doi.org/10.1007/398\\_2020\\_54](https://doi.org/10.1007/398_2020_54)
- Grube, A., Donaldson, D., Kiely, T., Wu, L. (2011). Pesticide Industry Sales and Usage. US EPA. <http://news.agropages.com/UserFiles/Report/Pdf/20120708204254971d.pdf>
- Guardian (2015) Hi-tech agriculture is freeing the farmer from his fields. 20<sup>th</sup> October 2015. <https://www.theguardian.com/environment/2015/oct/20/hi-tech-agriculture-is-freeing-farmer-from-his-fields>
- Guerrero Schimpf, M., Milesi, M. M., Ingaramo, P. I., Luque, E. H., & Varayoud, J. (2017). Neonatal exposure to a glyphosate based herbicide alters the development of the rat uterus. *Toxicology*, 376, 2–14. <https://doi.org/10.1016/j.tox.2016.06.004>
- Gunatilake, S., Seneff, S., & Orlando, L. (2019). Glyphosate's Synergistic Toxicity in Combination with Other Factors as a Cause of Chronic Kidney Disease of Unknown Origin. *International Journal of Environmental Research and Public Health*, 16(15), 2734. <https://doi.org/10.3390/ijerph16152734>
- Gurian-Sherman, D. (2009). *Failure to Yield - Evaluating the Performance of Genetically Engineered Crops*. Cambridge, MA: Union of Concerned Scientists.
- Harker, K. N., Mallory-Smith, C., Maxwell, B. D., Mortensen, D. A., & Smith, R. G. (2017). Another view. *Weed Science*, 65(2), 203–205. <https://doi.org/10.1017/wsc.2016.30>
- Hartzler, R. G. (2010). Reduction in common milkweed (*Asclepias syriaca*) occurrence in Iowa cropland from 1999 to 2009. *Crop Protection*, 29(12), 1542–1544. <http://doi.org/10.1016/j.cropro.2010.07.018>
- Hayhoe, S. J., Neill, C., Porder, S., Mchorney, R., Lefebvre, P., Coe, M. T., ... Krusche, A. V. (2011). Conversion to soy on the Amazonian agricultural frontier increases streamflow without affecting stormflow dynamics. *Global Change Biology*, 17(5), 1821–1833. <http://doi.org/10.1111/j.1365-2486.2011.02392.x>
- Heap, I. (2021). The International Herbicide-Resistant Weed Database. Online. Internet. Friday, August 27, 2021 . Available [www.weedscience.org](http://www.weedscience.org)
- Heap, I., & Duke, S. O. (2018). Overview of glyphosate-resistant weeds worldwide. *Pest Management Science*, 74(5), 1040–1049. <https://doi.org/10.1002/ps.4760>
- Heinemann, J. A., Massaro, M., Coray, D. S., Agapito-Tenfen, S. Z., & Wen, J. D. (2014a). Sustainability and innovation in staple crop production in the US Midwest. *International Journal of Agricultural Sustainability*, 12(1), 71–88. <http://doi.org/10.1080/14735903.2013.806408>

- Heinemann, J. A., Massaro, M., Coray, D. S., Agapito-Tenfen, S. Z. (2014b). Reply to comment on sustainability and innovation in staple crop production in the US Midwest. *International Journal of Agricultural Sustainability*, 12(4), 387-390. doi: 10.1080/14735903.2014.939843
- Hendlin, Y. H., Arcuri, A., Lepenies, R., & Hüesker, F. (2020). Like Oil and Water: The Politics of (Not) Assessing Glyphosate Concentrations in Aquatic Ecosystems. *European Journal of Risk Regulation*, 11(3), 539–564. <https://doi.org/10.1017/err.2020.65>
- Herek, J. S., Vargas, L., Rinas Trindade, S. A., Rutkoski, C. F., Macagnan, N., Hartmann, P. A., & Hartmann, M. T. (2021). Genotoxic effects of glyphosate on *Physalaemus* tadpoles. *Environmental Toxicology and Pharmacology*, 81, 103516. <https://doi.org/10.1016/j.etap.2020.103516>
- Herrera-Valdés, R., Almaguer-López, M. A., Orantes-Navarro, C. M., López-Marín, L., Brizuela-Díaz, E. G., Bayarre-Vea, H., Silva-Ayçaguer, L. C., Orellana de Figueroa, P., Smith-González, M., Chávez-Muñoz, Y., & Bacallao-Méndez, R. (2019). Epidemic of Chronic Kidney Disease of Nontraditional Etiology in El Salvador: Integrated Health Sector Action and South-South Cooperation. *MEDICC Review*, 21(4), 46–52. <https://doi.org/10.37757/MR2019.V21.N4.8>
- Hertel, R., Gibhardt, J., Martienssen, M., Kuhn, R., & Commichau, F. M. (2021). Molecular mechanisms underlying glyphosate resistance in bacteria. *Environmental Microbiology*, 23(6), 2891–2905. <https://doi.org/10.1111/1462-2920.15534>
- Hilbeck, A., Lebrecht, T., Vogel, R., Heinemann, J. A., & Binimelis, R. (2013). Farmer's choice of seeds in four EU countries under different levels of GM crop adoption. *Environmental Sciences Europe*, 25(1), 12. <http://doi.org/10.1186/2190-4715-25-12>
- Hoppin, J. A., Umbach, D. M., London, S. J., Henneberger, P. K., Kullman, G. J., Alavanja, M. C. R., & Sandler, D. P. (2008). Pesticides and atopic and nonatopic asthma among farm women in the Agricultural Health Study. *American Journal of Respiratory and Critical Care Medicine*, 177(1), 11–18. <http://doi.org/10.1164/rccm.200706-821OC>
- Hoppin, J. A., Umbach, D. M., Long, S., London, S. J., Henneberger, P. K., Blair, A., ... Sandler, D. P. (2016). Pesticides Are Associated with Allergic and Non-Allergic Wheeze among Male Farmers. *Environmental Health Perspectives*. <http://doi.org/10.1289/EHP315>
- Horticulture Week (2015) Ladybird Targeted spot sprayer offers reduced herbicide usage. 18<sup>th</sup> September 2015. <http://www.hortweek.com/ladybird-targeted-spot-sprayer-offers-reduced-herbicide-usage/products-kit/article/1364028>
- House of Commons Science and Technology Committee (2016) EU regulation of the life sciences. First Report of Session 2016–17. <http://www.publications.parliament.uk/pa/cm201617/cmselect/cmstech/158/158.pdf>
- Howard, P. H. (2009). Visualizing Consolidation in the Global Seed Industry: 1996–2008. *Sustainability*, 1(4), 1266–1287. <http://doi.org/10.3390/su1041266>
- Howe, C. M., Berrill, M., Pauli, B. D., Helbing, C. C., Werry, K., & Veldhoen, N. (2004). Toxicity of glyphosate-based pesticides to four North American frog species. *Environmental Toxicology and Chemistry*, 23(8), 1928–1938. <http://doi.org/10.1897/03-71>
- Hurley, L. (2022, June 21). U.S. Supreme Court rejects Bayer bid to nix Roundup weedkiller suits. *Reuters*. <https://www.reuters.com/legal/government/us-supreme-court-rejects-bayer-bid-nix-roundup-weedkiller-suits-2022-06-21/>
- IARC. (2018). *DDT, Lindane, and 2,4-D*. IARC Monographs on the Evaluation of Carcinogenic Risks to Humans, Lyon. Retrieved January 12, 2022, from <https://publications.iarc.fr/Book-And-Report-Series/Iarc-Monographs-On-The-Identification-Of-Carcinogenic-Hazards-To-Humans/DDT-Lindane-And-2-4-D-2016>
- IFOAM (2002). Position on genetic engineering and genetically modified organisms. <http://www.ifoam.org/sites/default/files/page/files/ifoam-ge-position.pdf>. Accessed 3 December 2014.
- Ingaramo, P. I., Varayoud, J., Milesi, M. M., Guerrero Schimpf, M., Alarcón, R., Muñoz-de-Toro, M., & Luque, E. H. (2017). Neonatal exposure to a glyphosate-based herbicide alters uterine decidualization in rats. *Reproductive Toxicology*, 73, 87–95. <https://doi.org/10.1016/j.reprotox.2017.07.022>
- Ingaramo, P., Alarcón, R., Muñoz-de-Toro, M., & Luque, E. H. (2020). Are glyphosate and glyphosate-based herbicides endocrine disruptors that alter female fertility? *Molecular and Cellular Endocrinology*, 110934. <https://doi.org/10.1016/j.mce.2020.110934>
- Ingaramo, P. I., Alarcón, R., Cagliaris, M. L., Varayoud, J., Muñoz-de-Toro, M., & Luque, E. H. (2022). Altered uterine angiogenesis in rats treated with a glyphosate-based herbicide. *Environmental Pollution*, 296, 118729. <https://doi.org/10.1016/j.envpol.2021.118729>
- ISAAA (2019a) ISAAA Brief 55-2019: Executive Summary | ISAAA.org. (2019). <https://www.isaaa.org/resources/publications/briefs/55/executivesummary/default.asp>
- Islam, F., Wang, J., Farooq, M. A., Khan, M. S. S., Xu, L., Zhu, J., Zhao, M., Muñoz, S., Li, Q. X., & Zhou, W. (2018). Potential impact of the herbicide 2,4-dichlorophenoxyacetic acid on human and ecosystems. *Environment International*, 111, 332–351. <https://doi.org/10.1016/j.envint.2017.10.020>

- Iturburu, F. G., Calderon, G., Amé, M. V., & Menone, M. L. (2019). Ecological Risk Assessment (ERA) of pesticides from freshwater ecosystems in the Pampas region of Argentina: Legacy and current use chemicals contribution. *Science of The Total Environment*, 691, 476–482. <https://doi.org/10.1016/j.scitotenv.2019.07.044>
- Janssens, L., & Stoks, R. (2017). Stronger effects of Roundup than its active ingredient glyphosate in damselfly larvae. *Aquatic Toxicology (Amsterdam, Netherlands)*, 193, 210–216. <https://doi.org/10.1016/j.aquatox.2017.10.028>
- Jaworski, E. G. (1972). Mode of action of N-phosphonomethylglycine. Inhibition of aromatic amino acid biosynthesis. *Journal of Agricultural and Food Chemistry*, 20(6), 1195–1198. <http://doi.org/10.1021/jf60184a057>
- Jayasumana C, Gunatilake S, Senanayake P (2014). Glyphosate, Hard Water and Nephrotoxic Metals: Are They the Culprits Behind the Epidemic of Chronic Kidney Disease of Unknown Etiology in Sri Lanka? *International Journal of Environmental Research and Public Health*, 11(2), 2125–2147. <http://doi.org/10.3390/ijerph110202125>
- Jayasumana C, Gunatilake S, Siribaddana S (2015) Simultaneous exposure to multiple heavy metals and glyphosate may contribute to Sri Lankan agricultural nephropathy. *BMC Nephrology*, 16, 103. <http://doi.org/10.1186/s12882-015-0109-2>
- Jia, R., Hou, Y., Feng, W., Li, B., & Zhu, J. (2022). Alterations at biochemical, proteomic and transcriptomic levels in liver of tilapia (*Oreochromis niloticus*) under chronic exposure to environmentally relevant level of glyphosate. *Chemosphere*, 294, 133818. <https://doi.org/10.1016/j.chemosphere.2022.133818>
- Johal, G. S. & Rahe, J. E. (1984). Effect of Soilborne Plant-Pathogenic Fungi on the Herbicidal Action of Glyphosate on Bean Seedlings. *Phytopathology*, 74(8), 950. <http://doi.org/10.1094/Phyto-74-950>
- Johal, G. S., & Huber, D. M. (2009). Glyphosate effects on diseases of plants. *European Journal of Agronomy*, 31(3), 144–152. <http://doi.org/10.1016/j.eja.2009.04.004>
- Kaboli Kafshgiri, S., Farkhondeh, T., & Miri-Moghaddam, E. (2021). Glyphosate effects on the female reproductive systems: A systematic review. *Reviews on Environmental Health*. <https://doi.org/10.1515/reveh-2021-0029>
- Kalofiri, P., Balias, G., & Tekos, F. (2021). The EU endocrine disruptors' regulation and the glyphosate controversy. *Toxicology Reports*, 8, 1193–1199. <https://doi.org/10.1016/j.toxrep.2021.05.013>
- King, C. A., Purcell, L. C., & Vories, E. D. (2001). Plant Growth and Nitrogenase Activity of Glyphosate-Tolerant Soybean in Response to Foliar Glyphosate Applications. *Agronomy Journal*, 93(1), 179. <http://doi.org/10.2134/agronj2001.931179x>
- Kissane, Z., & Shephard, J. M. (2017). The rise of glyphosate and new opportunities for biosentinel early-warning studies. *Conservation Biology*, 31(6), 1293–1300. <https://doi.org/10.1111/cobi.12955>
- Kremer, R. J., & Means, N. E. (2009). Glyphosate and glyphosate-resistant crop interactions with rhizosphere microorganisms. *European Journal of Agronomy*, 31(3), 153–161. <http://doi.org/10.1016/j.eja.2009.06.004>
- Krüger, M., Shehata, A. A., Schrödl, W., & Rodloff, A. (2013). Glyphosate suppresses the antagonistic effect of *Enterococcus* spp. on *Clostridium botulinum*. *Anaerobe*, 20, 74–78. <http://doi.org/10.1016/j.anaerobe.2013.01.005>
- Krüger, M., Philipp Schlendorn, Wieland Schrödl, Hans-Wolfgang Hoppe, Walburga Lutz, & Shehata, A. A. (2014a). Detection of Glyphosate Residues in Animals and Humans. *Journal of Environmental & Analytical Toxicology*, 04(02). <http://doi.org/10.4172/2161-0525.1000210>
- Krzysko-Lupicka, T., & Sudol, T. (2008). Interactions between glyphosate and autochthonous soil fungi surviving in aqueous solution of glyphosate. *Chemosphere*, 71(7), 1386–1391. <http://doi.org/10.1016/j.chemosphere.2007.11.006>
- Kumar, V., Liu, R., Boyer, G., & Stahlman, P. W. (2019). Confirmation of 2,4-D resistance and identification of multiple resistance in a Kansas Palmer amaranth (*Amaranthus palmeri*) population. *Pest Management Science*, 75(11), 2925–2933. <https://doi.org/10.1002/ps.5400>
- Kuphal K (2017) A shift away from GMO. Lakefield Standard. 20<sup>th</sup> February 2017. <http://www.lakefieldstandard.com/Stories/Story.cfm?SID=64527>
- Kurenbach, B., Marjoshi, D., Amábile-Cuevas, C. F., Ferguson, G. C., Godsoe, W., Gibson, P., & Heinemann, J. A. (2015). Sublethal Exposure to Commercial Formulations of the Herbicides Dicamba, 2,4-Dichlorophenoxyacetic Acid, and Glyphosate Cause Changes in Antibiotic Susceptibility in *Escherichia coli* and *Salmonella enterica* serovar Typhimurium. *mBio*, 6(2), e00009–15. <http://doi.org/10.1128/mBio.00009-15>
- Lapegna, P. (2013). The Expansion of Transgenic Soybeans and the Killing of Indigenous Peasants in Argentina. *ResearchGate*, 8(2), 291–308.
- Lapegna, P. (2016). Genetically modified soybeans, agrochemical exposure, and everyday forms of peasant collaboration in Argentina. *The Journal of Peasant Studies*, 43(2), 517–536. <http://doi.org/10.1080/03066150.2015.1041519>
- Larson, R. L., Hill, A. L., Fenwick, A., Kniss, A. R., Hanson, L. E., & Miller, S. D. (2006). Influence of glyphosate on *Rhizoctonia* and *Fusarium* root rot in sugar beet. *Pest Management Science*, 62(12), 1182–1192. <http://doi.org/10.1002/ps.1297>

- Le Du-Carrée, J., Boukhari, R., Cachot, J., Cabon, J., Louboutin, L., Morin, T., & Danion, M. (2021). Generational effects of a chronic exposure to a low environmentally relevant concentration of glyphosate on rainbow trout, *Oncorhynchus mykiss*. *Science of The Total Environment*, 801, 149462. <https://doi.org/10.1016/j.scitotenv.2021.149462>
- Le Du-Carrée, J., Cabon, J., Louboutin, L., Morin, T., & Danion, M. (2022). Changes in defense capacity to infectious hematopoietic necrosis virus (IHNV) in rainbow trout intergenerationally exposed to glyphosate. *Fish & Shellfish Immunology*, 122, 67–70. <https://doi.org/10.1016/j.fsi.2021.12.021>
- Leguizamón, A. (2016). Disappearing nature? Agribusiness, biotechnology and distance in Argentine soybean production. *The Journal of Peasant Studies*, 43(2), 313–330. <https://doi.org/10.1080/03066150.2016.1140647>
- Lerro, C. C., Hofmann, J. N., Andreotti, G., Koutros, S., Parks, C. G., Blair, A., Albert, P. S., Lubin, J. H., Sandler, D. P., & Beane Freeman, L. E. (2020). Dicamba use and cancer incidence in the agricultural health study: An updated analysis. *International Journal of Epidemiology*. <https://doi.org/10.1093/ije/dyaa066>
- Lesueur, C., Pirrotte, P., Pathak, K. V., Manservigi, F., Mandrioli, D., Belpoggi, F., Panzacchi, S., Li, Q., Barrett, E. S., Nguyen, R. H. N., Sathyanarayana, S., Swan, S. H., & Chen, J. (2021). Maternal urinary levels of glyphosate during pregnancy and anogenital distance in newborns in a US multicenter pregnancy cohort. *Environmental Pollution*, 280, 117002. <https://doi.org/10.1016/j.envpol.2021.117002>
- Liao, H., Li, X., Yang, Q., Bai, Y., Cui, P., Wen, C., Liu, C., Chen, Z., Tang, J., Che, J., Yu, Z., Geisen, S., Zhou, S., Friman, V.-P., & Zhu, Y.-G. (2021). Herbicide Selection Promotes Antibiotic Resistance in Soil Microbiomes. *Molecular Biology and Evolution*, 38(6), 2337–2350. <https://doi.org/10.1093/molbev/msab029>
- Little, A. (2022, July 13). Analysis | We Must Learn to Love Genetically Modified Crops. *Washington Post*. [https://www.washingtonpost.com/business/energy/we-must-learn-to-love-genetically-modified-crops/2022/07/12/be6f947a-01eb-11ed-8beb-2b4e481b1500\\_story.html](https://www.washingtonpost.com/business/energy/we-must-learn-to-love-genetically-modified-crops/2022/07/12/be6f947a-01eb-11ed-8beb-2b4e481b1500_story.html)
- Liu, Z., Shangguan, Y., Zhu, P., Sultan, Y., Feng, Y., Li, X., & Ma, J. (2022b). Developmental toxicity of glyphosate on embryo-larval zebrafish (*Danio rerio*). *Ecotoxicology and Environmental Safety*, 236, 113493. <https://doi.org/10.1016/j.ecoenv.2022.113493>
- Longhi, F., & Bianchi, S. (2020). Soy, glyphosate and human health. Some evidence in the Argentinian Dry Chaco Region (1990-2012). *Revista Geográfica de América Central*, 65, 145–174. <https://doi.org/10.15359/rgac.65-2.6>
- Lorenz, V., Pacini, G., Luque, E. H., Varayoud, J., & Milesi, M. M. (2020). Perinatal exposure to glyphosate or a glyphosate-based formulation disrupts hormonal and uterine milieu during the receptive state in rats. *Food and Chemical Toxicology: An International Journal Published for the British Industrial Biological Research Association*, 143, 111560. <https://doi.org/10.1016/j.fct.2020.111560>
- Louie, F., Jacobs, N. F. B., Yang, L. G. L., Park, C., Monnot, A. D., & Bandara, S. B. (2021). A comparative evaluation of dietary exposure to glyphosate resulting from recommended U.S. diets. *Food and Chemical Toxicology*, 158, 112670. <https://doi.org/10.1016/j.fct.2021.112670>
- Lozano-Kasten, F., Sierra-Díaz, E., Chavez, H. G., Peregrina Lucano, A. A., Cremades, R., & Pinto, E. S. (2021). Seasonal Urinary Levels of Glyphosate in Children From Agricultural Communities. *Dose-Response*, 19(4), 15593258211053184. <https://doi.org/10.1177/15593258211053184>
- Lu, G.-H., Hua, X.-M., Liang, L., Wen, Z.-L., Du, M.-H., Meng, F.-F., Pang, Y.-J., Qi, J.-L., Tang, C.-Y., & Yang, Y.-H. (2018). Identification of Major Rhizobacterial Taxa Affected by a Glyphosate-Tolerant Soybean Line via Shotgun Metagenomic Approach. *Genes*, 9(4), 214. <https://doi.org/10.3390/genes9040214>
- Lu, H., Lu, J., & He, L. (2019). Modeling and estimation of pollen-mediated gene flow at the landscape scale. *Ecological Indicators*, 106, 105500. <https://doi.org/10.1016/j.ecolind.2019.105500>
- Lupi, L., Bedmar, F., Puricelli, M., Marino, D., Aparicio, V. C., Wunderlin, D., & Miglioranza, K. S. B. (2019). Glyphosate runoff and its occurrence in rainwater and subsurface soil in the nearby area of agricultural fields in Argentina. *Chemosphere*, 225, 906–914. <https://doi.org/10.1016/j.chemosphere.2019.03.090>
- Lutri, V. F., Matteoda, E., Blarasin, M., Aparicio, V., Giacobone, D., Maldonado, L., Becher Quinodoz, F., Cabrera, A., & Giuliano Albo, J. (2020). Hydrogeological features affecting spatial distribution of glyphosate and AMPA in groundwater and surface water in an agroecosystem. Córdoba, Argentina. *Science of The Total Environment*, 711, 134557. <https://doi.org/10.1016/j.scitotenv.2019.134557>
- Macedo, M. N., Coe, M. T., DeFries, R., Uriarte, M., Brando, P. M., Neill, C., & Walker, W. S. (2013). Land-use-driven stream warming in southeastern Amazonia. *Phil. Trans. R. Soc. B*, 368(1619), 20120153. <http://doi.org/10.1098/rstb.2012.0153>
- Maggi, F., la Cecilia, D., Tang, F. H. M., & McBratney, A. (2020). The global environmental hazard of glyphosate use. *Science of The Total Environment*, 717, 137167. <https://doi.org/10.1016/j.scitotenv.2020.137167>
- Majewski, M. S., Coupe, R. H., Foreman, W. T., & Capel, P. D. (2014). Pesticides in Mississippi air and rain: a comparison between 1995 and 2007. *Environmental Toxicology and Chemistry / SETAC*, 33(6), 1283–1293. <http://doi.org/10.1002/etc.2550>

- Malcolm, S. B. (2018). Anthropogenic Impacts on Mortality and Population Viability of the Monarch Butterfly. *Annual Review of Entomology*, 63(1), 277–302. <https://doi.org/10.1146/annurev-ento-020117-043241>
- Mamy, L., Barriuso, E., & Gabrielle, B. (2016). Glyphosate fate in soils when arriving in plant residues. *Chemosphere*, 154, 425–433. <https://doi.org/10.1016/j.chemosphere.2016.03.104>
- Mann, R. M., & Bidwell, J. R. (1999). The Toxicity of Glyphosate and Several Glyphosate Formulations to Four Species of Southwestern Australian Frogs. *Archives of Environmental Contamination and Toxicology*, 36(2), 193–199. <http://doi.org/10.1007/s002449900460>
- Marc, J., Mulner-Lorillon, O., Boulben, S., Hureau, D., Durand, G., & Bellé, R. (2002). Pesticide Roundup provokes cell division dysfunction at the level of CDK1/cyclin B activation. *Chemical Research in Toxicology*, 15(3), 326–331.
- Marino, M., Mele, E., Viggiano, A., Nori, S. L., Meccariello, R., & Santoro, A. (2021). Pleiotropic Outcomes of Glyphosate Exposure: From Organ Damage to Effects on Inflammation, Cancer, Reproduction and Development. *International Journal of Molecular Sciences*, 22(22), 12606. <https://doi.org/10.3390/ijms222212606>
- Martinelli, L. A., Naylor, R., Vitousek, P. M., & Moutinho, P. (2010). Agriculture in Brazil: impacts, costs, and opportunities for a sustainable future. *Current Opinion in Environmental Sustainability*, 2(5–6), 431–438. <http://doi.org/10.1016/j.cosust.2010.09.008>
- Martinez, D. A., Loening, U. E., & Graham, M. C. (2018). Impacts of glyphosate-based herbicides on disease resistance and health of crops: A review. *Environmental Sciences Europe*, 30(1), 2. <https://doi.org/10.1186/s12302-018-0131-7>
- Martins-Gomes, C., Silva, T. L., Andreani, T., & Silva, A. M. (2022). Glyphosate vs. Glyphosate-Based Herbicides Exposure: A Review on Their Toxicity. *Journal of Xenobiotics*, 12(1), 21–40. <https://doi.org/10.3390/jox12010003>
- Mascarenhas, M., & Busch, L. (2006). Seeds of Change: Intellectual Property Rights, Genetically Modified Soybeans and Seed Saving in the United States. *Sociologia Ruralis*, 46(2), 122–138. <http://doi.org/10.1111/j.1467-9523.2006.00406.x>
- MASIPAG (2013). Socio-economic Impacts of Genetically Modified Corn In the Philippines. <http://masipag.org/wp-content/uploads/2013/05/MASIPAG-Book-low-res.pdf>
- Matozzo, V., Fabrello, J., & Marin, M. G. (2020). The Effects of Glyphosate and Its Commercial Formulations to Marine Invertebrates: A Review. *Journal of Marine Science and Engineering*, 8(6), 399. <https://doi.org/10.3390/jmse8060399>
- McKay, B., & Colque, G. (2016). Bolivia's soy complex: the development of “productive exclusion.” *The Journal of Peasant Studies*, 43(2), 583–610. <http://doi.org/10.1080/03066150.2015.1053875>
- Medalie, L., Baker, N. T., Shoda, M. E., Stone, W. W., Meyer, M. T., Stets, E. G., & Wilson, M. (2020). Influence of land use and region on glyphosate and aminomethylphosphonic acid in streams in the USA. *Science of The Total Environment*, 707, 136008. <https://doi.org/10.1016/j.scitotenv.2019.136008>
- Mesnage, R., & Antoniou, M. N. (2018). Ignoring Adjuvant Toxicity Falsifies the Safety Profile of Commercial Pesticides. *Frontiers in Public Health*, 5. <https://www.frontiersin.org/article/10.3389/fpubh.2017.00361>
- Mesnage, R., Christian Moesch, Le Grand, R., Guillaume, L., Vendomois, J. S., Gress, S., & Séralini, G.-E. (2012). Glyphosate Exposure in a Farmer's Family. *Journal of Environmental Protection*, 03(09), 1001–1003. <http://doi.org/10.4236/jep.2012.39115>
- Mesnage, R., Bernay, B., & Séralini, G.-E. (2013). Ethoxylated adjuvants of glyphosate-based herbicides are active principles of human cell toxicity. *Toxicology*, 313(2–3), 122–128. <http://doi.org/10.1016/j.tox.2012.09.006>
- Mesnage, R., Defarge, N., Vendomois, J. S., Séralini, G.-E. (2014). Major Pesticides Are More Toxic to Human Cells Than Their Declared Active Principles. *BioMed Research International*, 2014, e179691. <http://doi.org/10.1155/2014/179691>
- Mesnage, R., Benbrook, C., & Antoniou, M. N. (2019). Insight into the confusion over surfactant co-formulants in glyphosate-based herbicides. *Food and Chemical Toxicology*, 128, 137–145. <https://doi.org/10.1016/j.fct.2019.03.053>
- Milesi, M. M., Lorenz, V., Durando, M., Rossetti, M. F., & Varayoud, J. (2021). Glyphosate Herbicide: Reproductive Outcomes and Multigenerational Effects. *Frontiers in Endocrinology*, 12. <https://www.frontiersin.org/article/10.3389/fendo.2021.672532>
- Mitchell, D. G., Chapman, P. M., & Long, T. J. (1987). Acute toxicity of Roundup® and Rodeo® herbicides to rainbow trout, chinook, and coho salmon. *Bulletin of Environmental Contamination and Toxicology*, 39(6), 1028–1035. <http://doi.org/10.1007/BF01689594>
- Mohammadi, K., Sani, M. A., Safaei, P., Rahmani, J., Molaei-Aghae, E., & Jafari, S. M. (2021). A systematic review and meta-analysis of the impacts of glyphosate on the reproductive hormones. *Environmental Science and Pollution Research*. <https://doi.org/10.1007/s11356-021-16145-x>
- Mohapatra, S., Kumar, R., Sundaray, J. K., Patnaik, S. T., Mishra, C. S. K., & Rather, M. A. (2021). Structural damage in liver, gonads, and reduction in spawning performance and alteration in the haematological parameter of

- Anabas testudineus by glyphosate- a herbicide. *Aquaculture Research*, 52(3), 1150–1159. <https://doi.org/10.1111/are.14973>
- Monsanto. (2019, June 27). *Petition for the Determination of Nonregulated Status for Dicamba, Glufosinate, Quizalofop and 2,4-Dichlorophenoxyacetic Acid Tolerant MON 87429 Maize with Tissue-Specific Glyphosate Tolerance Facilitation the Production of Hybrid Maize Seed*. [https://www.aphis.usda.gov/brs/aphisdocs/19\\_31601p.pdf](https://www.aphis.usda.gov/brs/aphisdocs/19_31601p.pdf)
- Montiel-León, J. M., Munoz, G., Vo Duy, S., Do, D. T., Vaudreuil, M.-A., Goeury, K., Guillemette, F., Amyot, M., & Sauvé, S. (2019). Widespread occurrence and spatial distribution of glyphosate, atrazine, and neonicotinoids pesticides in the St. Lawrence and tributary rivers. *Environmental Pollution*, 250, 29–39. <https://doi.org/10.1016/j.envpol.2019.03.125>
- Moore, L. J., Fuentes, L., Rodgers Jr., J. H., Bowerman, W. W., Yarrow, G. K., Chao, W. Y., & Bridges Jr., W. C. (2012). Relative toxicity of the components of the original formulation of Roundup® to five North American anurans. *Ecotoxicology and Environmental Safety*, 78, 128–133. <http://doi.org/10.1016/j.ecoenv.2011.11.025>
- Moorman, T. B., Becerril, J. M., Lydon, J., & Duke, S. O. (1992). Production of hydroxybenzoic acids by Bradyrhizobium japonicum strains after treatment with glyphosate. *Journal of Agricultural and Food Chemistry*, 40(2), 289–293. <http://doi.org/10.1021/jf00014a025>
- Mordor Intelligence. (n.d.). Non-GMO Foods Market | 2022—27 | Industry Share, Size, Growth—Mordor Intelligence. Retrieved 8 June 2022, from <https://www.mordorintelligence.com/industry-reports/non-gmo-foods-market>
- Mortensen, D. A., Egan, J. F., Maxwell, B. D., Ryan, M. R., & Smith, R. G. (2012). Navigating a Critical Juncture for Sustainable Weed Management. *BioScience*, 62(1), 75–84. <http://doi.org/10.1525/bio.2012.62.1.12>
- Moutinho, M. F., de Almeida, E. A., Espíndola, E. L. G., Daam, M. A., & Schiesari, L. (2020). Herbicides employed in sugarcane plantations have lethal and sublethal effects to larval Boana pardalis (Amphibia, Hylidae). *Ecotoxicology*, 29(7), 1043–1051. <https://doi.org/10.1007/s10646-020-02226-z>
- Muñoz, J. P., Bleak, T. C., & Calaf, G. M. (2021). Glyphosate and the key characteristics of an endocrine disruptor: A review. *Chemosphere*, 270, 128619. <https://doi.org/10.1016/j.chemosphere.2020.128619>
- Murschell, T., & Farmer, D. K. (2019). Real-Time Measurement of Herbicides in the Atmosphere: A Case Study of MCPA and 2,4-D during Field Application. *Toxics*, 7(3), 40. <https://doi.org/10.3390/toxics7030040>
- Myers, J. P., Antoniou, M. N., Blumberg, B., Carroll, L., Colborn, T., Everett, L. G., ... Benbrook, C. M. (2016). Concerns over use of glyphosate-based herbicides and risks associated with exposures: a consensus statement. *Environmental Health*, 15(1). <http://doi.org/10.1186/s12940-016-0117-0>
- Nandula, V. K. (2019). Herbicide Resistance Traits in Maize and Soybean: Current Status and Future Outlook. *Plants*, 8(9), 337. <https://doi.org/10.3390/plants8090337>
- National Family Farm Coalition v. USEPA, No. 17–70196 (United States Court of Appeals for the Ninth Circuit June 3, 2020). <https://www.courthousenews.com/wp-content/uploads/2020/06/Dicamba.pdf>
- Neill, C., Coe, M. T., Riskin, S. H., Krusche, A. V., Eisenbeer, H., Macedo, M. N., ... Deegan, L. A. (2013). Watershed responses to Amazon soya bean cropland expansion and intensification. *Phil. Trans. R. Soc. B*, 368(1619), 20120425. <http://doi.org/10.1098/rstb.2012.0425>
- Newman, M. M., Hoilett, N., Lorenz, N., Dick, R. P., Liles, M. R., Ramsier, C., & Kloepper, J. W. (2016). Glyphosate effects on soil rhizosphere-associated bacterial communities. *Science of The Total Environment*, 543, Part A, 155–160. <http://doi.org/10.1016/j.scitotenv.2015.11.008>
- Nolan, E., & Santos, P. (2012). The Contribution of Genetic Modification to Changes in Corn Yield in the United States. *American Journal of Agricultural Economics*, aas069. <http://doi.org/10.1093/ajae/aas069>
- Nomura, H., Hamada, R., Wada, K., Saito, I., Nishihara, N., Kitahara, Y., Watanabe, S., Nakane, K., Nagata, C., Kondo, T., Kamijima, M., & Ueyama, J. (2022). Temporal trend and cross-sectional characterization of urinary concentrations of glyphosate in Japanese children from 2006 to 2015. *International Journal of Hygiene and Environmental Health*, 242, 113963. <https://doi.org/10.1016/j.ijheh.2022.113963>
- Non-GMO Project (2014). Non-GMO Month 2014 a Resounding Success. <http://www.nongmoproject.org/2014/11/14/non-gmo-month-2014-a-resounding-success/>
- Non-GMO Project (2015). Non-GMO Month 2015 Sets New Records, Generates Further Awareness. Living Non-GMO. <http://livingnongmo.org/2015/11/06/wrap-up-non-gmo-month-2015/>
- Non-GMO Project (2022) Product Verification. <https://www.nongmoproject.org/product-verification/>
- Nye, M., Hoilett, N., Ramsier, C., Renz, P., & P. Dick, R. (2014). Microbial Community Structure in Soils Amended With Glyphosate-tolerant Soybean Residue. *Applied Ecology and Environmental Sciences*, 2(3), 74–81. <http://doi.org/10.12691/aees-2-3-1>
- Odetti, L. M., López González, E. C., Romito, M. L., Simoniello, M. F., & Poletta, G. L. (2020). Genotoxicity and oxidative stress in Caiman latirostris hatchlings exposed to pesticide formulations and their mixtures during incubation period. *Ecotoxicology and Environmental Safety*, 193, 110312. <https://doi.org/10.1016/j.ecoenv.2020.110312>

- Oliva, A., Biasatti, R., Cloquell, S., González, C., Olego, S., & Gelin, A. (2008). Is there any relationship between rural environmental factors and reproductive health in the Pampa Humeda in Argentina? *Cadernos de Saúde Pública*, 24(4), 785–792. <http://doi.org/10.1590/S0102-311X2008000400008>
- Oliveira, N. P., Moi, G. P., Atanaka-Santos, M., Silva, A. M. C., & Pignati, W. A. (2014). Malformações congênitas em municípios de grande utilização de agrotóxicos em Mato Grosso, Brasil. *Ciência & Saúde Coletiva*, 19(10), 4123–4130. <http://doi.org/10.1590/1413-812320141910.08512014>
- Oliveira, G., & Hecht, S. (2016). Sacred groves, sacrifice zones and soy production: globalization, intensification and neo-nature in South America. *The Journal of Peasant Studies*, 43(2), 251–285. <http://doi.org/10.1080/03066150.2016.1146705>
- Oliver, M. (2020, June 4). Broadacre spot spraying kits: All you need to know. *Farmers Weekly*. <https://www.fwi.co.uk/machinery/technology/broadacre-spot-spraying-kits-all-you-need-to-know>
- Ortega, E., Cavalett, O., Bonifácio, R., & Watanabe, M. (2005). Brazilian Soybean Production: Emery Analysis With an Expanded Scope. *Bulletin of Science, Technology and Society*, 25(4), 323–334. <http://doi.org/10.1177/0270467605278367>
- Owagboriaye, F. O., Dedeke, G. A., Ademolu, K. O., Olujimi, O. O., Ashidi, J. S., & Adeyinka, A. A. (2017). Reproductive toxicity of Roundup herbicide exposure in male albino rat. *Experimental and Toxicologic Pathology*, 69(7), 461–468. <https://doi.org/10.1016/j.etp.2017.04.007>
- Paixão, F. (2020, November 18). *Transgenic HB4 wheat approved in Argentina has negative side effects*. Brasil de Fato. <https://www.brasildefato.com.br/2020/11/18/transgenic-hb4-wheat-approved-in-argentina-generates-negative-side-effects>
- PAN Asia and Pacific (2008) Monograph on Glyphosate. [http://www.panap.net/sites/default/files/monograph\\_glyphosate.pdf](http://www.panap.net/sites/default/files/monograph_glyphosate.pdf)
- PAN Asia and Pacific (2012) Monogram on Glyphosate: Addendum 2012. [http://www.panap.net/sites/default/files/monograph\\_glyphosate-addendum-2012.pdf](http://www.panap.net/sites/default/files/monograph_glyphosate-addendum-2012.pdf)
- Pannell, D.J., Tillie, P., Rodríguez-Cerezo, E., Ervin, D., Frisvold, G.B., 2017. Herbicide Resistance: Economic and Environmental Challenges. *AgBioForum* 19, 5.
- Parvez, S., Gerona, R. R., Proctor, C., Friesen, M., Ashby, J. L., Reiter, J. L., Lui, Z., & Winchester, P. D. (2018). Glyphosate exposure in pregnancy and shortened gestational length: A prospective Indiana birth cohort study. *Environmental Health*, 17, 23. <https://doi.org/10.1186/s12940-018-0367-0>
- Paull, J. (2018). Genetically Modified Organisms (GMOs) as Invasive Species. *Journal of Environment Protection and Sustainable Development*, 4(3), 31–37.
- Pereira, J. L., Antunes, S. C., Castro, B. B., Marques, C. R., Gonçalves, A. M. M., Gonçalves, F., & Pereira, R. (2009). Toxicity evaluation of three pesticides on non-target aquatic and soil organisms: commercial formulation versus active ingredient. *Ecotoxicology (London, England)*, 18(4), 455–463. <http://doi.org/10.1007/s10646-009-0300-y>
- Pereira, J. L., Lopes, M. C., Parish, J. B., Silva, A. A., & Picanço, M. C. (2018a). Impact of RR Soybeans and Glyphosate on the Community of Soil Surface Arthropods. *Planta Daninha*, 36. <https://doi.org/10.1590/S0100-83582018360100071>
- Pereira, J. L., Pereira, R. R., Resende-Silva, G. A., Jakelaitis, A., Silva, A. A., & Picanço, M. C. (2020). Glyphosate Impact on Arthropods Associated to Roundup Ready and Conventional Soybean (*Glycine max* L.). *Planta Daninha*, 38. <https://doi.org/10.1590/S0100-83582020380100047>
- Pérez, G. L., Vera, M. S., & Miranda, L. A. (2012). Effects of Herbicide Glyphosate and Glyphosate-Based Formulations on Aquatic Ecosystems. Retrieved from <http://cdn.intechweb.org/pdfs/12592.pdf>
- Perkins, P. J., Boermans, H. J., & Stephenson, G. R. (2000). Toxicity of glyphosate and triclopyr using the frog embryo teratogenesis assay—Xenopus. *Environmental Toxicology and Chemistry*, 19(4), 940–945. <http://doi.org/10.1002/etc.5620190422>
- Peters, A. (2021, January 9). This farming robot zaps weeds with precision lasers. *FastCompany*. <https://www.fastcompany.com/90670773/this-farming-robot-zaps-weeds-with-precision-lasers>
- Phélinas, P., & Choumert, J. (2017). Is GM Soybean Cultivation in Argentina Sustainable? *World Development*, 99, 452–462. <https://doi.org/10.1016/j.worlddev.2017.05.033>
- Pignati, W. A., Machado, J. M. H., & Cabral, J. F. (2007). Acidente rural ampliado: o caso das “chuvas” de agrotóxicos sobre a cidade de Lucas do Rio Verde - MT. *Ciência & Saúde Coletiva*, 12(1), 105–114. <http://doi.org/10.1590/S1413-81232007000100014>
- Piñeyro-Nelson, A., Van Heerwaarden, J., Perales, H. R., Serratos-Hernández, J. A., Rangel, A., Hufford, M. B., Gepts, P., Garay-Arroyo, A., Rivera-Bustamente, R., Alvarez-Buylla, E. R. (2009). Transgenes in Mexican maize: molecular evidence and methodological considerations for GMO detection in landrace populations. *Molecular Ecology*, 18(4), 750–761. <http://doi.org/10.1111/j.1365-294X.2008.03993.x>
- Pleasant, J. M., & Oberhauser, K. S. (2012). Milkweed loss in agricultural fields because of herbicide use: effect on the monarch butterfly population: Herbicide use and monarch butterflies. *Insect Conservation and Diversity*, 6(2), 135–144. <http://doi.org/10.1111/j.1752-4598.2012.00196.x>

- Pleasants, J. (2017). Milkweed restoration in the Midwest for monarch butterfly recovery: Estimates of milkweeds lost, milkweeds remaining and milkweeds that must be added to increase the monarch population. *Insect Conservation and Diversity*, 10(1), 42–53. <https://doi.org/10.1111/icad.12198>
- Pleasants, J. M., Williams, E. H., Brower, L. P., Oberhauser, K. S., & Taylor, O. R. (2016). Conclusion of No Decline in Summer Monarch Population Not Supported. *Annals of the Entomological Society of America*, sav115. <http://doi.org/10.1093/aesa/sav115>
- Pleasants, J. M., Zalucki, M. P., Oberhauser, K. S., Brower, L. P., Taylor, O. R., & Thogmartin, W. E. (2017). Interpreting surveys to estimate the size of the monarch butterfly population: Pitfalls and prospects. *PLOS ONE*, 12(7), e0181245. <https://doi.org/10.1371/journal.pone.0181245>
- Pompermaier, A., Varela, A. C. C., Mozzato, M. T., Soares, S. M., Fortuna, M., Alves, C., Tamagno, W. A., & Barcellos, L. J. G. (2022). Impaired initial development and behavior in zebrafish exposed to environmentally relevant concentrations of widely used pesticides. *Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology*, 257, 109328. <https://doi.org/10.1016/j.cbpc.2022.109328>
- Portier, C. J. (2020). A comprehensive analysis of the animal carcinogenicity data for glyphosate from chronic exposure rodent carcinogenicity studies. *Environmental Health*, 19(1), 18. <https://doi.org/10.1186/s12940-020-00574-1>
- Powles, S. B., & Preston, C. (2006). Evolved Glyphosate Resistance in Plants: Biochemical and Genetic Basis of Resistance<sup>1</sup>. *Weed Technology*, 20(2), 282–289. <http://doi.org/10.1614/WT-04-142R.1>
- Powelson, D. S., Stirling, C. M., Jat, M. L., Gerard, B. G., Palm, C. A., Sanchez, P. A., & Cassman, K. G. (2014). Limited potential of no-till agriculture for climate change mitigation. *Nature Climate Change*, 4(8), 678–683. <http://doi.org/10.1038/nclimate2292>
- Pratt, S. (2016a, January 8). Herbicide mixes tackle resistant weeds. *The Western Producer*. <https://www.producer.com/2016/01/herbicide-mixes-tackle-resistant-weeds/> [accessed: 9 April 2020]
- Price, B., & Cotter, J. (2014). The GM Contamination Register: a review of recorded contamination incidents associated with genetically modified organisms (GMOs), 1997–2013. *International Journal of Food Contamination*, 1(1), 5. <http://doi.org/10.1186/s40550-014-0005-8>
- Pu, Y., Yang, J., Chang, L., Qu, Y., Wang, S., Zhang, K., Xiong, Z., Zhang, J., Tan, Y., Wang, X., Fujita, Y., Ishima, T., Wang, D., Hwang, S. H., Hammock, B. D., & Hashimoto, K. (2020). Maternal glyphosate exposure causes autism-like behaviors in offspring through increased expression of soluble epoxide hydrolase. *Proceedings of the National Academy of Sciences of the United States of America*, 117(21), 11753–11759. <https://doi.org/10.1073/pnas.1922287117>
- Pu, Y., Ma, L., Shan, J., Wan, X., Hammock, B. D., & Hashimoto, K. (2021). Autism-like Behaviors in Male Juvenile Offspring after Maternal Glyphosate Exposure. *Clinical Psychopharmacology and Neuroscience*, 19(3), 554–558. <https://doi.org/10.9758/cpn.2021.19.3.554>
- Quartz (2015) This robot kills weeds, and could end the need for herbicides on farms. 18<sup>th</sup> November 2015. <http://qz.com/553383/this-robot-kills-weeds-and-could-end-the-need-for-herbicides-on-farms/>
- Quist, D., & Chapela, I. H. (2001). Transgenic DNA introgressed into traditional maize landraces in Oaxaca, Mexico. *Nature*, 414(6863), 541–543. <http://doi.org/10.1038/35107068>
- Ranum, P., Peña-Rosas, J. P., & Garcia-Casal, M. N. (2014). Global maize production, utilization, and consumption. *Annals of the New York Academy of Sciences*, 1312, 105–112. <http://doi.org/10.1111/nyas.12396>
- Ramirez Haberkon, N. B., Aparicio, V. C., & Mendez, M. J. (2021). First evidence of glyphosate and aminomethylphosphonic acid (AMPA) in the respirable dust (PM10) emitted from unpaved rural roads of Argentina. *Science of The Total Environment*, 773, 145055. <https://doi.org/10.1016/j.scitotenv.2021.145055>
- Ramsdorf, W., Bordin, E., Munhoz, R., Panicio, P., & Freitas, A. (2021). Transgenerational Effects of Environmentally Relevant Concentrations of Atrazine and Glyphosate Herbicides, Isolated and in Mixture, to Freshwater Microcrustacean *Daphnia Magna*. <https://doi.org/10.21203/rs.3.rs-790734/v1>
- Raoult, D., Hadjadj, L., Baron, S. A., & Rolain, J.-M. (2021). Role of glyphosate in the emergence of antimicrobial resistance in bacteria? *Journal of Antimicrobial Chemotherapy*, 76(7), 1655–1657. <https://doi.org/10.1093/jac/dkab102>
- Redo, D., Aide, T. M., & Clark, M. L. (2013). Vegetation change in Brazil's dryland ecoregions and the relationship to crop production and environmental factors: Cerrado, Caatinga, and Mato Grosso, 2001–2009. *Journal of Land Use Science*, 8(2), 123–153. <http://doi.org/10.1080/1747423X.2012.667448>
- Relyea, R. A. (2005a). The Impact of Insecticides and Herbicides on the Biodiversity and Productivity of Aquatic Communities. *Ecological Applications*, 15(2), 618–627. <http://doi.org/10.1890/03-5342>
- Relyea, R. A. (2005b). The lethal impact of roundup on aquatic and terrestrial amphibians. *Ecological Applications*, 15(4), 1118–1124. <http://doi.org/10.1890/04-1291>
- Relyea, R. A. (2005c). The Lethal Impacts of Roundup and Predatory Stress on Six Species of North American Tadpoles. *Archives of Environmental Contamination and Toxicology*, 48(3), 351–357. <http://doi.org/10.1007/s00244-004-0086-0>

- Reuters (2016c) USDA confirms unapproved GMO wheat found in Washington state. 29 July 2016. <http://www.reuters.com/article/us-wheat-washington-gmo-idUSKCN10920K>
- Riaño, C., Ortiz-Ruiz, M., Pinto-Sánchez, N. R., & Gómez-Ramírez, E. (2020). Effect of glyphosate (Roundup Active®) on liver of tadpoles of the colombian endemic frog *Dendropsophus molitor* (amphibia: Anura). *Chemosphere*, 250, 126287. <https://doi.org/10.1016/j.chemosphere.2020.126287>
- Richard, S., Moslemi, S., Sipahutar, H., Benachour, N., & Seralini, G.-E. (2005). Differential Effects of Glyphosate and Roundup on Human Placental Cells and Aromatase. *Environmental Health Perspectives*, 113(6), 716–720.
- Rigotto, R. M., Vasconcelos, D. P. e, Rocha, M. M., Rigotto, R. M., Vasconcelos, D. P. e, & Rocha, M. M. (2014). Pesticide use in Brazil and problems for public health. *Cadernos de Saúde Pública*, 30(7), 1360–1362. <http://doi.org/10.1590/0102-311XPE020714>
- Rissoli, R. Z., Abdalla, F. C., Costa, M. J., Rantin, F. T., McKenzie, D. J., & Kalinin, A. L. (2016). Effects of glyphosate and the glyphosate based herbicides Roundup Original® and Roundup Transorb® on respiratory morphophysiology of bullfrog tadpoles. *Chemosphere*, 156, 37–44. <https://doi.org/10.1016/j.chemosphere.2016.04.083>
- Rodrigues, N. R., & de Souza, A. P. F. (2018). Occurrence of glyphosate and AMPA residues in soy-based infant formula sold in Brazil. *Food Additives & Contaminants: Part A*, 35(4), 724–731. <https://doi.org/10.1080/19440049.2017.1419286>
- Rodrigues, N. R., Souza, A. P. F. de, Morais, P. P. P., Braga, D. P. V., Crivellari, A. C., Favoretto, L. R. G., & Berger, G. U. (2020). Residues of glyphosate and aminomethylphosphonic acid (AMPA) in genetically modified glyphosate tolerant soybean, corn and cotton crops. *Ciência Rural*, 51. <https://doi.org/10.1590/0103-8478cr20190244>
- Rodríguez, E. M., Medesani, D. A., Canosa, I. S., & Avigliano, L. (2021). The Effect of Glyphosate on the Reproduction of Estuarine Crabs: *Neohelice granulata* as a Study Model. *Frontiers in Endocrinology*, 12. <https://www.frontiersin.org/article/10.3389/fendo.2021.643168>
- Romano, R. M., Romano, M. A., Bernardi, M. M., Furtado, P. V., & Oliveira, C. A. (2010). Prepubertal exposure to commercial formulation of the herbicide glyphosate alters testosterone levels and testicular morphology. *Archives of Toxicology*, 84(4), 309–317. <http://doi.org/10.1007/s00204-009-0494-z>
- Roseboro, K. (2008): Finding non-GMO soybean seed becoming more difficult: fewer breeding programs for non-GMO soybeans are reducing supplies despite strong demand. The Organic and Non-GMO Report. [http://www.non-gmoreport.com/articles/jul08/non-gmo\\_soybean\\_seed.php](http://www.non-gmoreport.com/articles/jul08/non-gmo_soybean_seed.php)
- Roseboro, K. (2012). Scientist says biotech companies encouraging GMO-herbicide treadmill. The Organic and Non-GMO Report. <http://www.non-gmoreport.com/articles/april2012/scientist-biotech-gmo-herbicide.php>
- Roseboro, K. (2015b). More Farmers Predicted to Go Non-GMO and Organic in 2015. Organic Connections. <http://organicconnectmag.com/more-farmers-predicted-to-go-non-gmo-and-organic-in-2015/>
- Ruuskanen, S., Rainio, M. J., Uusitalo, M., Saikkonen, K., & Helander, M. (2020a). Effects of parental exposure to glyphosate-based herbicides on embryonic development and oxidative status: A long-term experiment in a bird model. *Scientific Reports*, 10(1), 6349. <https://doi.org/10.1038/s41598-020-63365-1>
- Ruuskanen, S., Rainio, M. J., Gomez-Gallego, C., Selenius, O., Salminen, S., Collado, M. C., Saikkonen, K., Saloniemi, I., & Helander, M. (2020b). Glyphosate-based herbicides influence antioxidants, reproductive hormones and gut microbiome but not reproduction: A long-term experiment in an avian model. *Environmental Pollution*, 115108. <https://doi.org/10.1016/j.envpol.2020.115108>
- Ruuskanen, S., Rainio, M. J., Kuosmanen, V., Laihonon, M., Saikkonen, K., Saloniemi, I., & Helander, M. (2020c). Female Preference and Adverse Developmental Effects of Glyphosate-Based Herbicides on Ecologically Relevant Traits in Japanese Quails. *Environmental Science & Technology*, 54(2), 1128–1135. <https://doi.org/10.1021/acs.est.9b07331>
- Ryan, C. D. & Smyth, S. J. (2012). Economic Implications of Low-level Presence in a Zero-Tolerance European Import Market: The Case of Canadian Triffid Flax. *AgBioForum*, 15(1), 21-30. <http://www.agbioforum.org/v15n1/v15n1a03-ryan.htm>
- Sabio y García, C. A., Vera, M. S., Vinocur, A., Graziano, M., Miranda, C., & Pizarro, H. N. (2022). Rethinking the term “glyphosate effect” through the evaluation of different glyphosate-based herbicide effects over aquatic microbial communities. *Environmental Pollution*, 292, 118382. <https://doi.org/10.1016/j.envpol.2021.118382>
- Sánchez, J. A. A., Barros, D. M., de los Angeles Bistoni, M., Ballesteros, M. L., Roggio, M. A., & Martins, C. D. G. M. (2021). Glyphosate-based herbicides affect behavioural patterns of the livebearer *Jenynsia multidentata*. *Environmental Science and Pollution Research*, 28(23), 29958–29970. <https://doi.org/10.1007/s11356-020-11958-8>
- Sanogo, S., Yang, X. B., & Scherm, H. (2000). Effects of Herbicides on *Fusarium solani* f. sp. *glycines* and Development of Sudden Death Syndrome in Glyphosate-Tolerant Soybean. *Phytopathology*, 90(1), 57–66. <http://doi.org/10.1094/PHYTO.2000.90.1.57>
- Sanogo, S., Yang, X. B., & Lundeen, P. (2001). Field Response of Glyphosate-Tolerant Soybean to Herbicides and Sudden Death Syndrome. *Plant Disease*, 85(7), 773–779. <http://doi.org/10.1094/PDIS.2001.85.7.773>

- Santadino, M., Coviella, C., & Momo, F. (2014). Glyphosate Sublethal Effects on the Population Dynamics of the Earthworm *Eisenia fetida* (Savigny, 1826). *Water, Air, & Soil Pollution*, 225(12), 1–8. <http://doi.org/10.1007/s11270-014-2207-3>
- Santos-Silva, T., Ribeiro, R. I. M. A., Memorian, S. N. A. (In Thomé, R. G., & Santos, H. (2021). *Combinatorial Effects of Organophosphate (Temephos), Glyphosate (Roundup) and Sodium Dodecil Sulfate (Sds) on Zebrafish (Danio Rerio) Gills: A Morphometric and Histological Study*. <https://doi.org/10.21203/rs.3.rs-185080/v1>
- Savitz, D. A., Arbuckle, T., Kaczor, D., & Curtis, K. M. (1997). Male Pesticide Exposure and Pregnancy Outcome. *American Journal of Epidemiology*, 146(12), 1025–1036.
- Schaefer, A. & Carter, A. (2015). GMO trade in a world of fragmented consumer preferences and needs. International Centre for Trade and Sustainable Development. <http://www.ictsd.org/bridges-news/biores/news/gmo-trade-in-a-world-of-fragmented-consumer-preferences-and-needs>
- Schiesari, L., & Grillitsch, B. (2011). Pesticides meet megadiversity in the expansion of biofuel crops. *Frontiers in Ecology and the Environment*, 9(4), 215–221. <http://doi.org/10.1890/090139>
- Schmidt, M., López, V. T., Tobias, M., Grinberg, E., & Merlinsky, G. (2022). Social and environmental conflicts caused by agrochemical use in Salta, Santiago del Estero and Santa Fe, Argentina. *Ciência & Saúde Coletiva*, 27, 1061–1072. <https://doi.org/10.1590/1413-81232022273.04852021>
- Schulz, A., Krüper, A., & Amrhein, N. (1985). Differential sensitivity of bacterial 5-enolpyruvylshikimate-3-phosphate synthases to the herbicide glyphosate. *FEMS Microbiology Letters*, 28(3), 297–301. <http://doi.org/10.1111/j.1574-6968.1985.tb00809.x>
- Schulz, R., Bub, S., Petschick, L. L., Stehle, S., & Wolfram, J. (2021). Applied pesticide toxicity shifts toward plants and invertebrates, even in GM crops. *Science*, 372(6537), 81–84. <https://doi.org/10.1126/science.abe1148>
- Serra, L., Estienne, A., Vasseur, C., Froment, P., & Dupont, J. (2021). Review: Mechanisms of Glyphosate and Glyphosate-Based Herbicides Action in Female and Male Fertility in Humans and Animal Models. *Cells*, 10(11), 3079. <https://doi.org/10.3390/cells10113079>
- Servizi, J. A., Gordon, R. W., & Martens, D. W. (1987). Acute toxicity of Garlon 4 and Roundup herbicides to Salmon, Daphnia, and trout. *Bulletin of Environmental Contamination and Toxicology*, 39(1), 15–22. <http://doi.org/10.1007/BF01691783>
- Shehata, A. A., Schrödl, W., Aldin, A. A., Hafez, H. M., & Krüger, M. (2013). The Effect of Glyphosate on Potential Pathogens and Beneficial Members of Poultry Microbiota In Vitro. *Current Microbiology*, 66(4), 350–358. <http://doi.org/10.1007/s00284-012-0277-2>
- Silva, J. F. S. da, Silva, A. M. C. da, Lima-Luz, L., Aydos, R. D., & Mattos, I. E. (2015). Correlação entre produção agrícola, variáveis clínicas-demográficas e câncer de próstata: um estudo ecológico. *Ciência & Saúde Coletiva*, 20(9), 2805–2812. <http://doi.org/10.1590/1413-81232015209.00582015>
- Silver, M. K., Fernandez, J., Tang, J., McDade, A., Sabino, J., Rosario, Z., V. élez V. C., Alshawabkeh, A., Cordero, J. F., & Meeker, J. D. (2021). Prenatal Exposure to Glyphosate and Its Environmental Degradate, Aminomethylphosphonic Acid (AMPA), and Preterm Birth: A Nested Case–Control Study in the PROTECT Cohort (Puerto Rico). *Environmental Health Perspectives*, 129(5), 057011. <https://doi.org/10.1289/EHP7295>
- Sineiro, C. C., & Berger, M. (2012). Citizens' Rights and Environmental Genocide. *Environmental Justice*, 5(2), 105–110. <http://doi.org/10.1089/env.2011.0011>
- Singh, S., Kumar, V., Datta, S., Wani, A. B., Dhanjal, D. S., Romero, R., & Singh, J. (2020). Glyphosate uptake, translocation, resistance emergence in crops, analytical monitoring, toxicity and degradation: A review. *Environmental Chemistry Letters*, 18(3), 663–702. <https://doi.org/10.1007/s10311-020-00969-z>
- Slaby, S., Titran, P., Marchand, G., Hanotel, J., Lescuyer, A., Leprière, A., Bodart, J.-F., Marin, M., & Lemiere, S. (2020). Effects of glyphosate and a commercial formulation Roundup® exposures on maturation of *Xenopus laevis* oocytes. *Environmental Science and Pollution Research*, 27(4), 3697–3705. <https://doi.org/10.1007/s11356-019-04596-2>
- Slager, R. E., Poole, J. A., LeVan, T. D., Sandler, D. P., Alavanja, M. C. R., & Hoppin, J. A. (2009). Rhinitis associated with pesticide exposure among commercial pesticide applicators in the Agricultural Health Study. *Occupational and Environmental Medicine*, 66(11), 718–724. <http://doi.org/10.1136/oem.2008.041798>
- Smyth, S., Khachatourians, G. G., Philips, P. W. B. (2002). Liabilities and economics of transgenic crops. *Nature Biotechnology*, vol. 20, 537–541.
- Snow, A. (2009). Unwanted transgenes re-discovered in Oaxacan maize. *Molecular Ecology*, 18(4), 569–571. <http://doi.org/10.1111/j.1365-294X.2008.04063.x>
- Sohn, S.-I., Pandian, S., Oh, Y.-J., Kang, H.-J., Ryu, T.-H., Cho, W.-S., Shin, E.-K., & Shin, K.-S. (2021). A Review of the Unintentional Release of Feral Genetically Modified Rapeseed into the Environment. *Biology*, 10(12), 1264. <https://doi.org/10.3390/biology10121264>
- Soltani, N., Oliveira, M. C., Alves, G. S., Werle, R., Norsworthy, J. K., Sprague, C. L., Young, B. G., Reynolds, D. B., Brown, A., & Sikkema, P. H. (2020). Off-target movement assessment of dicamba in North America. *Weed Technology*, 34(3), 318–330. <https://doi.org/10.1017/wet.2020.17>

- Southey, F. (2021, February 11). *Behind the push for GMO transparency on-pack: 'Non-GMO food labelling closes a loophole in EU legislation'*. Foodnavigator.Com. <https://www.foodnavigator.com/Article/2021/02/11/ENGA-pushes-for-non-GMO-food-labelling-across-Europe>
- Sprinkle, R. H., & Payne-Sturges, D. C. (2021). Mixture toxicity, cumulative risk, and environmental justice in United States federal policy, 1980–2016. *Environmental Health*, 20(1), 104. <https://doi.org/10.1186/s12940-021-00764-5>
- Stempel, J. (2022, June 17). U.S. EPA ordered to reassess glyphosate's impact on health, environment. *Reuters*. <https://www.reuters.com/business/environment/us-agency-ordered-reassess-glyphosates-impact-health-environment-2022-06-17/>
- Stenoien, C., Nail, K. R., Zalucki, J. M., Parry, H., Oberhauser, K. S., & Zalucki, M. P. (2018). Monarchs in decline: A collateral landscape-level effect of modern agriculture. *Insect Science*, 25(4), 528–541. <https://doi.org/10.1111/1744-7917.12404>
- Strandberg, B., Sørensen, P. B., Bruus, M., Bossi, R., Dupont, Y. L., Link, M., & Damgaard, C. F. (2021). Effects of glyphosate spray-drift on plant flowering. *Environmental Pollution*, 280, 116953. <https://doi.org/10.1016/j.envpol.2021.116953>
- Struger, J., Thompson, D., Staznik, B., Martin, P., McDaniel, T., & Marvin, C. (2008). Occurrence of Glyphosate in Surface Waters of Southern Ontario. *Bulletin of Environmental Contamination and Toxicology*, 80(4), 378–384. <http://doi.org/10.1007/s00128-008-9373-1>
- Suppa, A., Kvist, J., Li, X., Dhandapani, V., Almulla, H., Tian, A. Y., Kissane, S., Zhou, J., Perotti, A., Mangelson, H., Langford, K., Rossi, V., Brown, J. B., & Orsini, L. (2020). Roundup causes embryonic development failure and alters metabolic pathways and gut microbiota functionality in non-target species. *Microbiome*, 8(1), 170. <https://doi.org/10.1186/s40168-020-00943-5>
- Tan, S., Li, G., Liu, Z., Wang, H., Guo, X., & Xu, B. (2022). Effects of glyphosate exposure on honeybees. *Environmental Toxicology and Pharmacology*, 90, 103792. <https://doi.org/10.1016/j.etap.2021.103792>
- Tang, X., Sretenovic, S., Ren, Q., Jia, X., Li, M., Fan, T., Yin, D., Xiang, S., Guo, Y., Liu, L., Zheng, X., Qi, Y., & Zhang, Y. (2020). Plant Prime Editors Enable Precise Gene Editing in Rice Cells. *Molecular Plant*, 13(5), 667–670. <https://doi.org/10.1016/j.molp.2020.03.010>
- Taylor, O. R. J., Pleasants, J. M., Grundel, R., Pecoraro, S. D., Lovett, J. P., & Ryan, A. (2020). Evaluating the Migration Mortality Hypothesis Using Monarch Tagging Data. *Frontiers in Ecology and Evolution*, 0. <https://doi.org/10.3389/fevo.2020.00264>
- Thogmartin, W. E., Wiederholt, R., Oberhauser, K., Drum, R. G., Diffendorfer, J. E., Altizer, S., Taylor, O. R., Pleasants, J., Semmens, D., Semmens, B., Erickson, R., Libby, K., & Lopez-Hoffman, L. (2017). Monarch butterfly population decline in North America: Identifying the threatening processes. *Royal Society Open Science*, 4(9), 170760. <https://doi.org/10.1098/rsos.170760>
- Thongprakaisang, S., Thiantanawat, A., Rangkadilok, N., Suriyo, T., & Satayavivad, J. (2013). Glyphosate induces human breast cancer cells growth via estrogen receptors. *Food and Chemical Toxicology*, 59, 129–136. <http://doi.org/10.1016/j.fct.2013.05.057>
- Tillie, P., Rodríguez-Cerezo, E. (2015). Markets for non-Genetically Modified Identity-Preserved soybean in the EU. JRC Science and Policy Report, Europea Commission. <http://publications.jrc.ec.europa.eu/repository/bitstream/JRC95457/report.pdf>
- Tresnakova, N., Stara, A., & Velisek, J. (2021). Effects of Glyphosate and Its Metabolite AMPA on Aquatic Organisms. *Applied Sciences*, 11(19), 9004. <https://doi.org/10.3390/app11199004>
- Tscharntke, T., Clough, Y., Wanger, T. C., Jackson, L., Motzke, I., Perfecto, I., ... Whitbread, A. (2012). Global food security, biodiversity conservation and the future of agricultural intensification. *Biological Conservation*, 151(1), 53–59. <http://doi.org/10.1016/j.biocon.2012.01.068>
- Tsui, M. T. K., & Chu, L. M. (2003). Aquatic toxicity of glyphosate-based formulations: comparison between different organisms and the effects of environmental factors. *Chemosphere*, 52(7), 1189–1197. [http://doi.org/10.1016/S0045-6535\(03\)00306-0](http://doi.org/10.1016/S0045-6535(03)00306-0)
- Tsui, M. T. K., & Chu, L. M. (2004). Comparative Toxicity of Glyphosate-Based Herbicides: Aqueous and Sediment Porewater Exposures. *Archives of Environmental Contamination and Toxicology*, 46(3), 316–323. <http://doi.org/10.1007/s00244-003-2307-3>
- Twellman, B. (2021, November 29). *Non-GMO U.S. Soy Production Competes with High Commodity Prices*. U.S. Soy. <https://ussoy.org/non-gmo-u-s-soy-production-competes-with-high-commodity-prices/?persona=importers&pillar=exceptional-supply&region=americas&goal=inform-educate>
- Unglesbee, E. (2020b). *Dicamba Not Controlling Some Tennessee Palmer Amaranth Populations*. DTN Progressive Farmer. <https://www.dtnpf.com/agriculture/web/ag/crops/article/2020/02/27/dicamba-controlling-tennessee-palmer>
- Unglesbee, E. (2020f, June 4). *Dicamba Registrations Vacated—The Ninth Circuit Vacates Three Dicamba Registrations*. DTN Progressive Farmer. <https://www.dtnpf.com/agriculture/web/ag/crops/article/2020/06/04/ninth-circuit-vacates-three-dicamba>

- Unglesbee, E. (2020n, July 14). *The Future of Dicamba—Dicamba Use Faces Trio of Threats: Courts, Weeds and Farmer Fatigue*. DTN Progressive Farmer. <https://www.dtnpf.com/agriculture/web/ag/crops/article/2020/07/14/dicamba-use-faces-trio-threats-weeds>
- Unglesbee, E. (2021a, February 17). *Glufosinate-Resistant Palmer Amaranth Confirmed in Arkansas*. DTN Progressive Farmer. <https://www.dtnpf.com/agriculture/web/ag/crops/article/2021/02/17/glufosinate-resistant-palmer>
- Unglesbee, E. (2022a, March 29). *Court Asks EPA for Agency's Long-Term Dicamba Plan*. DTN Progressive Farmer. <https://www.dtnpf.com/agriculture/web/ag/crops/article/2022/03/29/court-asks-epa-agencys-long-term>
- Unglesbee, E. (2022b, July 1). *Court Asked to Kickstart Lawsuit to Vacate Dicamba Registrations*. DTN Progressive Farmer. <https://www.dtnpf.com/agriculture/web/ag/crops/article/2022/01/07/court-asked-kickstart-lawsuit-vacate>
- Upamaliika, S. W. A. M., Wannige, C. T., Vidanagamachchi, S. M., Gunasekara, S. C., Kolli, R. T., De Silva, P. M. C. S., Kulasiri, D., & Jayasundara, N. (2022). A review of molecular mechanisms linked to potential renal injury agents in tropical rural farming communities. *Environmental Toxicology and Pharmacology*, 92, 103850. <https://doi.org/10.1016/j.etap.2022.103850>
- USDA NASS (2015). 2012 Census of Agriculture. Organic Survey (2014). Volume 3, Special Studies, Part 4. [http://www.agcensus.usda.gov/Publications/2012/Online\\_Resources/Organics/ORGANICS.pdf](http://www.agcensus.usda.gov/Publications/2012/Online_Resources/Organics/ORGANICS.pdf)
- US EPA. (1998). *Pesticide Fact Sheet*. United States Environmental Protection Agency, Office of Prevention, Pesticides and Toxic Substances. [https://www3.epa.gov/pesticides/chem\\_search/reg\\_actions/registration/fs\\_PC-123000\\_15-Sep-98.pdf](https://www3.epa.gov/pesticides/chem_search/reg_actions/registration/fs_PC-123000_15-Sep-98.pdf)
- Van Bruggen, A. H. C., He, M. M., Shin, K., Mai, V., Jeong, K. C., Finckh, M. R., & Morris, J. G. (2018). Environmental and health effects of the herbicide glyphosate. *Science of The Total Environment*, 616–617, 255–268. <https://doi.org/10.1016/j.scitotenv.2017.10.309>
- Van Bruggen, A. H. C., Finckh, M. R., He, M., Ritsema, C. J., Harkes, P., Knuth, D., & Geissen, V. (2021). Indirect Effects of the Herbicide Glyphosate on Plant, Animal and Human Health Through its Effects on Microbial Communities. *Frontiers in Environmental Science*, 9. <https://www.frontiersin.org/article/10.3389/fenvs.2021.763917>
- Vanlaeys, A., Dubuisson, F., Seralini, G.-E., & Travert, C. (2018). Formulants of glyphosate-based herbicides have more deleterious impact than glyphosate on TM4 Sertoli cells. *Toxicology in Vitro*, 52, 14–22. <https://doi.org/10.1016/j.tiv.2018.01.002>
- Varayoud, J., Durando, M., Ramos, J. G., Milesi, M. M., Ingaramo, P. I., Muñoz-de-Toro, M., & Luque, E. H. (2017). Effects of a glyphosate-based herbicide on the uterus of adult ovariectomized rats. *Environmental Toxicology*, 32(4), 1191–1201. <https://doi.org/10.1002/tox.22316>
- Vázquez, M. B., Moreno, M. V., Amodeo, M. R., & Bianchinotti, M. V. (2021). Effects of glyphosate on soil fungal communities: A field study. *Revista Argentina de Microbiología*, 53(4), 349–358. <https://doi.org/10.1016/j.ram.2020.10.005>
- Vidal, O., & Rendón-Salinas, E. (2014). Dynamics and trends of overwintering colonies of the monarch butterfly in Mexico. *Biological Conservation*, 180, 165–175. <http://doi.org/10.1016/j.biocon.2014.09.041>
- Vieira, B. C., Luck, J. D., Amundsen, K. L., Werle, R., Gaines, T. A., & Kruger, G. R. (2020). Herbicide drift exposure leads to reduced herbicide sensitivity in *Amaranthus* spp. *Scientific Reports*, 10(1), 2146. <https://doi.org/10.1038/s41598-020-59126-9>
- Vila-Aiub, M. M., Balbi, M. C., Gundel, P. E., Ghersa, C. M., & Powles, S. B. (2007). Evolution of Glyphosate-Resistant Johnsongrass (*Sorghum halepense*) in Glyphosate-Resistant Soybean. *Weed Science*, 55(6), 566–571.
- Vila-Aiub, M. M., Vidal, R. A., Balbi, M. C., Gundel, P. E., Trucco, F., & Ghersa, C. M. (2008). Glyphosate-resistant weeds of South American cropping systems: an overview. *Pest Management Science*, 64(4), 366–371. <http://doi.org/10.1002/ps.1488>
- Viljoen, C. D., Koortzen, B. J., & Sreenivasan Tantuan, S. (2021). Determining the presence of glyphosate and glyphosate-tolerant events in maize and soybean food products in South Africa. *Food Additives & Contaminants: Part B*, 14(2), 91–97. <https://doi.org/10.1080/19393210.2021.1872713>
- Vincent, K., & Davidson, C. (2015). The toxicity of glyphosate alone and glyphosate-surfactant mixtures to western toad (*Anaxyrus boreas*) tadpoles. *Environmental Toxicology and Chemistry / SETAC*. <http://doi.org/10.1002/etc.3118>
- Vivian, R., Reis, A., Kálnay, P. A., Vargas, L., & Mariani, A. C. C. F. and F. (2013). Weed Management in Soybean — Issues and Practices. InTech. <http://www.intechopen.com/books/soybean-pest-resistance/weed-management-in-soybean-issues-and-practices>
- Walsh, L. P., McCormick, C., Martin, C., & Stocco, D. M. (2000). Roundup inhibits steroidogenesis by disrupting steroidogenic acute regulatory (StAR) protein expression. *Environmental Health Perspectives*, 108(8), 769–776.

- Walsh, M., Ouzman, J., Newman, P., Powles, S., & Llewellyn, R. (2017). High Levels of Adoption Indicate That Harvest Weed Seed Control Is Now an Established Weed Control Practice in Australian Cropping. *Weed Technology*, 31(3), 341–347. <https://doi.org/10.1017/wet.2017.9>
- Wegier, A., Piñeyro-Nelson, A., Alarcón, J., Gálvez-Mariscal, A., Alvarez-Buylla, E. R., & Piñero, D. (2011). Recent long-distance transgene flow into wild populations conforms to historical patterns of gene flow in cotton (*Gossypium hirsutum*) at its centre of origin. *Molecular Ecology*, 20(19), 4182–4194. <http://doi.org/10.1111/j.1365-294X.2011.05258.x>
- Where is Glyphosate Banned? (2022, March). Baum Hedlund Aristei & Goldman. <https://www.baumhedlundlaw.com/toxic-tort-law/monsanto-roundup-lawsuit/where-is-glyphosate-banned/>
- Xu, J., Smith, S., Smith, G., Wang, W., & Li, Y. (2019). Glyphosate contamination in grains and foods: An overview. *Food Control*, 106, 106710. <https://doi.org/10.1016/j.foodcont.2019.106710>
- Yang, M., Wen, Z., Fazal, A., Hua, X., Xu, X., Yin, T., Qi, J., Yang, R., Lu, G., Hong, Z., & Yang, Y. (2020). Impact of a G2-EPSPS & GAT Dual Transgenic Glyphosate-Resistant Soybean Line on the Soil Microbial Community under Field Conditions Affected by Glyphosate Application. *Microbes and Environments*, 35(4), ME20056. <https://doi.org/10.1264/jsme2.ME20056>
- Yannicari, M., Vázquez-García, J. G., Gómez-Lobato, M. E., Rojano-Delgado, A. M., Alves, P. L. da C. A., & De Prado, R. (2021). First Case of Glyphosate Resistance in *Bromus catharticus* Vahl.: Examination of Endowing Resistance Mechanisms. *Frontiers in Plant Science*, 12, 198. <https://doi.org/10.3389/fpls.2021.617945>
- Yield10 Bioscience. (2022, March 3). *Yield10 Bioscience Announces an Update on the Camelina Line E3902 Development Program for Producing Low-carbon Feedstock Oil for Renewable Diesel | Press Release*. <https://www.yield10bio.com/press/yield10-bioscience-announces-an-update-on-the-camelina-line-e3902-development-program-for-producing-low-carbon-feedstock-oil-for-renewable-diesel>
- Young, F., Ho, D., Glynn, D., Edwards, V. (2015). Endocrine disruption and cytotoxicity of glyphosate and roundup in human Jar cells *in vitro*. *Integrative Pharmacology, Toxicology and Genotoxicology*. Vol. 1(1): 12-19. doi: 10.15761/IPTG.1000104.
- Yu, Z., Lu, C., Hennessy, D. A., Feng, H., & Tian, H. (2020). Impacts of tillage practices on soil carbon stocks in the US corn-soybean cropping system during 1998 to 2016. *Environmental Research Letters*, 15(1), 014008. <https://doi.org/10.1088/1748-9326/ab6393>
- Zaller, J. G., Heigl, F., Ruess, L., & Grabmaier, A. (2014). Glyphosate herbicide affects belowground interactions between earthworms and symbiotic mycorrhizal fungi in a model ecosystem. *Scientific Reports*, 4. <http://doi.org/10.1038/srep05634>
- Zalucki, M. P., & Lammers, J. H. (2010). Dispersal and egg shortfall in Monarch butterflies: what happens when the matrix is cleaned up? *Ecological Entomology*, 35(1), 84–91. <http://doi.org/10.1111/j.1365-2311.2009.01160.x>
- Zhao, L., Zhang, J., Yang, L., Zhang, H., Zhang, Y., Gao, D., Jiang, H., Li, Y., Dong, H., Ma, T., Wang, X., Wu, M., Wang, A., Jin, Y., Yuan, Y., & Chen, H. (2021). Glyphosate exposure attenuates testosterone synthesis via NR1D1 inhibition of StAR expression in mouse Leydig cells. *Science of The Total Environment*, 785, 147323. <https://doi.org/10.1016/j.scitotenv.2021.147323>
- Zheng, W., Scott, J. W., Holm, N., & Machesky, M. L. (2018). *Occurrence and Fate of the Herbicide Glyphosate in Tile Drainage and Receiving Rivers in East Central Illinois*. Illinois Sustainable Technology Center. <http://hdl.handle.net/2142/101912>
- Zhou, X., Larson, J.A., Lambert, D.M., Roberts, R.K., English, B.C., Bryant, K.J., Mishra, A.K., Falconer, L.L., Hogan Jr., R.J., Johnson, J.L., & Reeves, J.M. (2015). Farmer experience with weed resistance to herbicides in cotton production. *AgBioForum*, 18(1), 114-125. <http://agbioforum.org/v18n1/v18n1a12-zhou.htm>
- Zhou, Z., Ni, X., Wu, Z., & Tang, J. (2022). Physiological and transcriptomic analyses reveal the threat of herbicides glufosinate and glyphosate to the scleractinian coral *Pocillopora damicornis*. *Ecotoxicology and Environmental Safety*, 229, 113074. <https://doi.org/10.1016/j.ecoenv.2021.113074>
- Zhu, L., Li, W., Zha, J., & Wang, Z. (2015). Dicamba affects sex steroid hormone level and mRNA expression of related genes in adult rare minnow (*Gobiocypris rarus*) at environmentally relevant concentrations. *Environmental Toxicology*, 30(6), 693–703. <https://doi.org/10.1002/tox.21947>
- Zilberman, D., Sexton, S. E., Marra, M. C., & Fernandez-Cornejo, J. (2010). The Economic Impact of Genetically Engineered Crops. *Choices*, 25(2). Retrieved from <http://ideas.repec.org/a/ags/aaeach/94769.htm>
- Zobiole, L. H. S., Bonini, E. A., Jr, R. S. de O., Kremer, R. J., & Ferrarese-Filho, O. (2010a). Glyphosate affects lignin content and amino acid production in glyphosate-resistant soybean. *Acta Physiologiae Plantarum*, 32(5), 831–837. <http://doi.org/10.1007/s11738-010-0467-0>
- Zobiole, L. H. S., Jr, R. S. de O., Huber, D. M., Constantin, J., Castro, C. de, Oliveira, F. A. de, & Jr, A. de O. (2010b). Glyphosate reduces shoot concentrations of mineral nutrients in glyphosate-resistant soybeans. *Plant and Soil*, 328(1-2), 57–69. <http://doi.org/10.1007/s11104-009-0081-3>

- Zobiolo, L. H. S., Oliveira, R. S., Visentainer, J. V., Kremer, R. J., Bellaloui, N., & Yamada, T. (2010c). Glyphosate Affects Seed Composition in Glyphosate-Resistant Soybean. *Journal of Agricultural and Food Chemistry*, *58*(7), 4517–4522. <http://doi.org/10.1021/jf904342t>
- Zobiolo, L. H. S., Kremer, R. J., Oliveira, R. S., & Constantin, J. (2011). Glyphosate affects micro-organisms in rhizospheres of glyphosate-resistant soybeans. *Journal of Applied Microbiology*, *110*(1), 118–127. <http://doi.org/10.1111/j.1365-2672.2010.04864.x>
- Zobiolo, L. H. S., Kremer, R. J., de Oliveira Jr., R. S., & Constantin, J. (2012). Glyphosate effects on photosynthesis, nutrient accumulation, and nodulation in glyphosate-resistant soybean. *Journal of Plant Nutrition and Soil Science*, *175*(2), 319–330. <http://doi.org/10.1002/jpln.201000434>